

POLLEN MORPHOLOGY AND TAXONOMY IN THE LOGANIACEAE

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Pollen Morphology

(W. Punt)

The Loganiaceae is a heterogeneous, eurypalynous family with colpate, colporate or porate pollen grains (Erdtman 1952). Some years ago Dr. Leeuwenberg, specialist in the taxonomy of African Loganiaceae, asked the senior author to undertake an investigation of the pollen grains of that family. Unfortunately that was impossible at the time because of other commitments. Later, however, a possibility presented itself for carrying out the investigation in connection with a sojourn at the Palynological Laboratory at Solna. I am much indebted to Professor Erdtman for the invitation to work at this Laboratory, for his approval of the subject, and for many discussions on pollen morphology. I am also much indebted to Dr. Leeuwenberg, Wageningen, and Dr. Leenhouts, Leiden for interesting, carefully determined plant material and for kind advice in taxonomic problems. I also want to express my thanks to all those in the Solna Laboratory, who kindly helped me in various ways during my visit. My work was supported by a grant from the U.S. Atomic Energy Commission to the Palynological Laboratory.

Materials and Methods

Herbarium material from the following botanical institutions was used: Botanisch Museum en Herbarium, Utrecht (U); Rijksherbarium, Leiden (L); Laboratorium voor Plantensystematiek en -geo-

grafie, Wageningen (WAG); Naturhistoriska Riksmuseet Stockholm (S). Some specimens from various herbaria were sent to Dr. Leeuwenberg at Wageningen. Besides, some slides from the sporothèque of the Palynological Laboratory Solna (SPL) were used.

The dry material was first boiled in water and then acetolysed (after washing with glacial acetic acid). Acetolysis was carried out in two ways: 1. "Macromethod" (Erdtman 1960). 2. "Micromethod" (Punt 1962). The latter method was used only when a very minute quantity of material was present. The specimens are marked "Mi" or "Ma" in Table 1 according to their treatment. The mounting medium was glycerine jelly. After mounting the slides were sealed off with paraffine wax.

Pollen grains of *Labordia tinifolia*, *Spigelia marilandica*, and *Anthocleista nobilis* were sectioned with a Spencer microtome (model 812) in the way described by Praglowski (in Erdtman 1957).

The pollen slides were examined with a Leitz Ortholux microscope (apochromatic objective 96 ×, A 1.32; eyepiece Periplan GF, 10 ×). The measurements were, as a rule, based on five grains only. The figures thus have no statistical significance. The photomicrographs were taken with a Leitz Ortholux microscope, objective C P1 Oel 160 ×, A 1.40. Ilford Chromatic, Panchromatic and sometimes Half Tone Chromatic plates were used. Yellow, orange or green filters were used. Original magnification usually 1500 ×.

The terminology used in this paper follows Erdtman (1943 and 1952) and Fægri and Iversen (1964). In some cases alternatives are given in brackets.

Pollen grains from all genera so far described were examined. In the following list the number of species examined is compared with the total number of species (according to Leenhouts, 1962):

Number of species examined		Total number of spp.	Number of species examined		Total number of spp.
1. <i>Antonia</i>	1	1	10. <i>Fagraea</i>	26	ca. 31
2. <i>Adenoplea</i>	1	ca. 2	11. <i>Gardneria</i>	3	5
3. <i>Adenoplusia</i>	2	ca. 2	12. <i>Gelsemium</i>	2	3
4. <i>Androya</i>	1	1	13. <i>Geniostoma</i>	3	ca. 30
5. <i>Bonyunia</i>	4	ca. 4	14. <i>Gomphostigma</i>	1	2
6. <i>Buddleia</i>	13	ca. 110	15. <i>Labordia</i>	4	ca. 20
7. <i>Cynoctonum</i>	3	6	16. <i>Logania</i>	5	ca. 20
8. <i>Desfontainia</i>	1	1	17. <i>Mitrasacme</i>	6	ca. 40
9. <i>Emorya</i>	1	1	18. <i>Mostuea</i>	4	8

Number of species examined	Total number of spp.	Number of species examined	Total number of spp.		
19. <i>Neuburgia</i>	4	ca. 10	25. <i>Retzia</i>	1	1
20. <i>Norrisia</i>	2	2	26. <i>Sanango</i>	1	1
21. <i>Nuxia</i>	6	ca. 20	27. <i>Spigelia</i>	3	ca. 50
22. <i>Peltanthera</i>	1	1	28. <i>Strychnos</i>	6	ca. 200
23. <i>Polypremum</i>	1	1	29. <i>Usteria</i>	1	1
24. <i>Potalia</i>	1	1	Gen. inc. sed.: <i>Plocosperma</i>	1	1

Pollen descriptions and comments

ANTONIEAE

ANTONIA-TYPE

Apertures. — Pollen grains 3-colporate. Colpi sunken. Colpus membrane smooth. Ora (endopori) circular to slightly lologate, as wide as colpi. Margins of ora thickened (costae).

Shape. — Equatorial view: grains more or less spheroidal (subprolate in *Bonyunia superba*). Polar view: grains circular.

Exine. — Sexine thicker than nexine. Tectum smooth, usually perforate. In *Bonyunia aquatica* the bacules (columellae) form a regular pattern (infrastrate-infrareticulate). In the other species of *Bonyunia* the bacules are inordinate.

<i>Antonia ovata</i>	18 × 19 μ	OSph	
—Pl. 1: 11–12			
<i>Bonyunia antoniifolia</i>	30 × 33 μ	OSph	
<i>B. aquatica</i> —Pl. 1: 13	26 × 26 μ	Sph	exine infra-rugulose, muri straight
<i>B. minor</i>	26 × 24 μ	PSph	
<i>B. superba</i>	28 × 24 μ	SP	bacula distinct, longer at poles
<i>Norrisia maior</i>	13 × 14 μ	OSph or Sph	
<i>N. malaccensis</i>	17 × 17.5 μ	OSph or Sph	
<i>Usteria guineensis</i>	22 × 22 μ	Sph	

Antonia, *Bonyunia*, *Norrisia* and *Usteria* have pollen grains of the Antonia-type. Its most striking character is the circular ora accentuated by costae. The exine characters are very similar to those in the Logania-type and the Spigelia-type. Distinct ora are also found in the Gelsemium-type. A smooth perforated tectum is also present in the Logania-, Spigelia-, Potalia- and Geniostoma-types.

Pollen grains of the Antonia-type differ markedly from those in the genus *Peltanthera*, which undoubtedly belong to the Buddleia

type. This fits with the opinion of Leenhouts (1962) who included *Peltanthera* in the Buddleieae whereas Hutchinson (1959) placed it in the Antoniaceae. The pollen grains of Oleaceae are entirely different.

Antonia (S. America), *Usteria* (Africa) and *Norrisia* (Malesia) show a close resemblance in their pollen. It is remarkable that these small and geographically widely separated genera are so much alike in their pollen grains. The pollen grains of *Bonyunia* no doubt also belong to this type but they differ slightly in their exine structure. *Bonyunia aquatica* has short inordinate lirae (rugulate, Fægri & Iversen, 1964). The other species of *Bonyunia* differ more or less in their bacules characters. The differences are so small that no attempt for a further division of the genus is made here.

Neither illustrations nor chromosome numbers are published.

BUDDLEIEAE

BUDDLEIA-TYPE

Buddleia subtype

Apertures. — Pollen grains 3-colporate, sometimes 4-colporate. Colpi sunken, ends frequently blunt. Colpus membrane smooth. Ora lalongate (endocolpi), small and indistinct, ends diffuse. Margins not marked by costae.

Shape. — Equatorial view: grains usually subprolate to spheroidal, sometimes oblate-spheroidal. Polar view: grains circular, rarely goniotreme (semiangular or subangular). Grains rather small (longest axis not exceeding 25 μ).

Exine. — Sexine thicker than nexine. Tectum smooth. Bacules (columellae) frequently indistinct.

<i>Adenoplea sinuata</i>	18 × 18 μ	Sph	Finely reticulate (D'Alleizette s.n.)
—	17 × 15.5 μ	PSph	Not-reticulate (Afzelius s.n.)
<i>Adenoplusia axillaris</i>	ca. 15 μ		Bad material
<i>A. uluguruensis</i>	16 × 17.5 μ	OSph	
<i>Androya decaryi</i>	20 × 18.5 μ	PSph	Ora indistinct
<i>Buddleia alpina</i>	18.5 × 18 μ	Sph or PSph	Pollen grains sometimes 4-colporate
<i>B. americana</i>	17 × 18 μ	OSph	Usually 4-colporate
<i>B. coriacea</i>	21 × 17.5 μ	SP	
<i>B. corrugata</i>	19 × 15.5 μ	SP	
<i>B. elegans</i>	21 × 21 μ	Sph	Slightly angular in polar view

<i>B. indica</i> —Pl. 1: 1–2	20 × 17.5 μ	SP or PSph	
<i>B. madagascariensis</i>	16 × 15 μ	PSph	
<i>B. marrubifolia</i>	17 × 15.5 μ	PSph	
<i>B. salvifolia</i>	16 × 13.5 μ	SP	
<i>B. stachyoides</i>	25 × 21 μ	SP	
<i>Nuxia capitata</i>	15 × 13 μ	SP	Ora indistinct and narrow
<i>N. congesta</i>	14 × 15.5 μ	OSph	
<i>N. keniensis</i> (syn. <i>N. congesta</i>)	17.5 × 15 μ	SP	
<i>N. oppositifolia</i> (syn. <i>N. autunesii</i>)	15.5 × 13 μ	SP	
<i>Peltanthera floribunda</i>	14 × 14 μ	Sph	
<i>Sanango durum</i>	15.5 × 16 μ	Sph or Osph	

Pollen grains of this subtype occur in most species of *Buddleia* (sect. *Chilianthus* excluded), most species of *Nuxia* and in some small genera close to *Buddleia*, viz., *Adenoplea*, *Adenoplusia*, *Androya*, *Peltanthera*, and *Sanango*.

Chilianthus subtype

Apertures. — Pollen grains 3-colporate. Colpi sunken, Colpus membrane smooth. Ora lalongate (endocolpi), small and indistinct, their ends diffuse. Margins without costae.

Shape. — Equatorial view: grains spheroidal to subprolate. Polar view: grains circular or somewhat goniotreme (semiangular).

Exine. — Sexine thicker than nexine. Grains reticulate. Lumina irregular, small.

<i>Buddleia dysophylla</i>	17 × 15 μ	Psph or SP	
<i>B. glomerata</i>	17 × 13 μ	SP	
<i>B. saligna</i> — Pl. 1: 3–5	16 × 14 μ	Psph or SP	
<i>Gomphostigma virgatum</i>	21 × 21 μ	Sph	(Pont 1718)
	23 × 25.5 μ	Osph	(Norlindh & Weimarck 4499)
<i>Nuxia floribunda</i> (syn. <i>N. polyantha</i>)	13.5 × 13.5 μ	Sph	
<i>N. verticillata</i>	15 × 14 μ	PSph	reticulum fine, transition to a perforate tectum

This subtype is found in several species of *Buddleia* (sect. *Chilianthus*), some species of *Nuxia* and in the small genus *Gomphostigma*. The *Chilianthus* subtype closely resembles the *Buddleia* subtype. The only difference is found in the exine. The pollen grains

in *Gomphostigma* differ from those in *Buddleia* and *Nuxia* in size only.

EMORYA-TYPE

Apertures. — Pollen grains 4-colporate (sometimes loxotreme). Colpi sunken. Colpus membrane smooth. Ora lalongate (endocolpi), small, indistinct, their ends diffuse. Margins without costae.

Shape. — Equatorial view: grains subprolate to prolate (sometimes depressed at the poles). Polar view: grains circular to slightly angular.

Exine. — Sexine thicker than nexine. Tectum smooth, perforate. Bacules (columellae) inordinate, comparatively long, sometimes placed in groups.

Emorya suaveolens — Pl. 1:6–10 42 × 32 μ SP-P

Only *Emorya* belongs to this type. It resembles the *Buddleia*-type in some characteristics (especially the ora). The number of colpi, however, is different and the bacules are always distinct.

The pollen grains of the *Buddleia* subtype show some slight variation in their morphological characters. There is some fluctuation in the shape, in the length of the colpi, and in the distinctness of the bacules.

The *Chilianthus* subtype differs from the *Buddleia* subtype only by the presence of a reticulum. In some species (e.g. *Nuxia verticillata*) the reticulum is so fine that it represents a transition to the non-reticulate *Buddleia* subtype. Therefore it seems better to unite these subtypes with the *Buddleia*-type.

The *Emorya*-type differs markedly from the *Buddleia*-type by its shape, by having four colpi, and by the distinctly visible bacules. Some species in the *Buddleia*-type also have four colpi (*B. alpina* and *B. americana*). The most important similarity between both types is in the ora which are small and indistinct. The notable difference in pollen grains between *Emorya* and *Buddleia* is in contrast with the overall resemblance between the two genera which made Leenhouts (1962) that they might be united.

The two types have little in common with other Loganiaceous types being considerably different in shape and apertures.

The pollen grains of *Peltanthera* and *Sanango* agree in all respects with the *Buddleia* subtype and do not share any character with the *Antonia*-type (*Antonieae*). The genera *Adenoplea*, *Adenoplusia*, *Androya*, and subgenus *Nicodemia* (*Buddleia*) also have pollen grains

which clearly belong to the *Buddleia* subtype. The relationship of *Nuxia* with *Buddleia* is also confirmed by the pollen morphological results.

There is some pollen morphological support to place the *Buddleieae* near the *Scrophulariaceae*. The resemblance between their pollen grains and those of *Calceolaria* is so striking that the latter genus cannot be excluded from the *Buddleia*-type. The pollen grains of the genera *Antirrhinum*, *Halleria*, *Linaria*, *Scrophularia*, and *Verbascum* also have several characters in common with the *Chilianthus* subtype. The grains of the genera *Alonsia*, *Aplosium*, *Castilleja*, *Digitalis*, *Hebe*, *Hebenstreitia* (*Selaginaceae*), *Hydrotriche*, and *Paulownia* are completely different. This selection of genera is arbitrary and the list is far from complete.

According to Leenhouts (1962) the *Buddleieae* form a transition between the *Loganiaceae* and the *Scrophulariaceae*; they are, however, much easier to separate from the latter than from the former family.

Reduction of the genus *Chilianthus* to *Buddleia* is not supported by pollenmorphological evidence. *Chilianthus* Burch. was united with *Buddleia* Linn. by Phillips (1946) who was followed by Verdoorn (1963). The following three species belong to this taxon: *Buddleia saligna* Willd. [syn. *Chilianthus arboreus* (Linn. f.) A.DC.], *Buddleia dysophylla* (Benth.) Radlk. [syn. *Ch. dysophyllus* (Benth.) A.DC.], *Buddleia glomerata* Wendl. f. [syn. *Ch. lobulatus* (Benth.) A.DC.]. One species, *Buddleia corrugata* (Benth.) Phillips, forms a transition between "*Chilianthus*" and *Buddleia*. Pollen grains of this species undoubtedly belong to the *Buddleia* subtype. On the other hand, some *Nuxia* species (*N. polyantha* and *N. verticillata*) also have pollen grains of the *Chilianthus* subtype. This is an indication that *Nuxia* and *Chilianthus* may be closely related. In fact, all *Chilianthus* species have at one time or another been included in *Nuxia*.

Illustrations

Erdtman 1952, fig. 144B, *Buddleia japonica* Hemsl.

Wang 1960, Tab. LXII, *Buddleia lindleyana* Forst.

Chromosome numbers (Gadella 1962, 1963, Moore 1947, 1960)

<i>Buddleia albiflora</i> Hemsl.	$2n = \text{ca. } 114$
<i>B. alternifolia</i> Maxim.	38
<i>B. americana</i> Linn.	76

<i>B. asiatica</i> Lour.	38
<i>B. brasiliensis</i> Jacq.	38
<i>B. candida</i> Dunn.	76
<i>B. colvilei</i> Hook. f. et Thoms.	ca. 300
<i>B. crispa</i> Benth.	38
<i>B. davidii</i> Franch.	76
<i>B. delavayi</i> Gagnep.	ca. 114
<i>B. fallowiana</i> Balf. f. et W. W. Smith	76
<i>B. jarreri</i> Balf. f. et W. W. Smith	38
<i>B. forestii</i> Diels	ca. 114
<i>B. globosa</i> Hope	38
<i>B. glandiflora</i> Champ. et Schl.	38
<i>B. hastata</i> Prain ex Marq.	38
<i>B. indica</i> Lam.	76
<i>B. japonica</i> Hemsl.	38
<i>B. limitanea</i> Smith	ca. 114
<i>B. lindleyana</i> Fort.	38
<i>B. madagascariensis</i> Lam.	38
<i>B. nappii</i> Lorenz	38
<i>B. nivea</i> Duthie	ca. 114
<i>B. paniculata</i> Wall.	38
<i>B. pterocaulis</i> Jacks.	ca. 228
<i>B. salvifolia</i> (Linn.) Lam.	38
<i>B. scordioides</i> H. B. K.	38
<i>B. stenostachya</i> Reh. et Wils.	ca. 114
<i>B. sterniana</i> Cotton	38
<i>Chilianthus arboreus</i> (Linn. f.) DC.	38
<i>Nuxia floribunda</i> Benth.	38

SPIGELIEAE (p. p.)

SPIGELIA-TYPE

Apertures. — Pollen grains 3-colpate or 3-colporate. Colpi wide, not sunken, rather short, and their ends sharp. Colpus membrane absent or present only at the ends of the colpi. Ora congruent with the colpi in *Spigelia*, nearly congruent in *Desfontainia* and *Polypremum*.

Shape. — Equatorial view: grains suboblate or oblate. Polar view: grains goniotreme (semiangular or triangular).

Exine. — Sexine thicker than nexine. Tectum smooth, perforate. Bacules (columellae) short and different in shape and size.

<i>Desfontainia spinosa</i> — Pl. 2: 6-7	39 × 45 μ	SO or O
<i>Polypremum procumbens</i>	24.5 × 29 μ	SO
<i>Spigelia anthelmia</i>	34 × 62 μ	O
<i>S. leiocarpa</i> — Pl. 2: 5	45 × 62 μ	O
<i>S. martiana</i>	29 × 43 μ	O

Spigelia, *Desfontainia* and *Polypreum* have pollen grains of this type. Colpus membranes are not present in the investigated species of *Spigelia*. In *Desfontainia* and *Polypreum*, however, a small part of the colpus membrane is present at the ends of the colpi. If there is no colpus membrane, we should speak of an endo-aperture. In the genera *Desfontainia* and *Polypreum*, where a small part of the colpus membrane is left, it seems reasonable to speak of colporate grains. In *Spigelia*, where the colpus membrane is completely wanting, it seems better to speak of colpate grains, although, strictly theoretically, these grains are also colporate. The most remarkable character of the *Spigelia*-type is the absence or, indistinctness of the ora. This character separates the type from all other types in the Loganiaceae. There is, however, a similarity in shape and exine structure with the *Logania*-type.

The pollen grains of *Desfontainia* fit completely in the *Spigelia*-type and do not show any resemblance with the *Potalia*-type. Inclusion of *Desfontainia* in the Loganiaceae is supported by this result.

The pollen grains of *Polypreum*, although less oblate in shape than the grains of *Spigelia* and *Desfontainia*, are placed in the same type because of the indistinct lologate ora.

The taxonomic position of *Desfontainia* is not clear. Solereder (1892), Klett (1924), and Leenhouts (1962) were doubtful with regard to its affinity. The two last named authors suggested a place near the *Potalieae*. Pollenmorphological characters are in favour of a place near *Spigelia* and *Polypreum*.

The pollen grains of the Rubiaceae-Hedyotideae, to which group Leenhouts (1962) believed *Polypreum* to be rather closely related, are different.

Pollen of *Mitrasacme elata* are similar in shape and apertures but differ in exine structure (reticulate).

MITRASACME ELATA-TYPE

Apertures. — Pollen grains 3-colporate. Colpi wide, not sunken, rather short, their ends sharp. Colpus membrane present only at the ends of the colpi. Ora lologate, indistinct.

Shape. — Equatorial view: grains suboblate or oblate. Polar view: grains goniotreme (angular).

Exine. — Sexine thicker than nexine. Grains reticulate. Muri simpli-baculate. Lumina 1–3 μ , smaller towards colpi and poles.

Mitrasacme elata var. *elata*

31 × 41 μ

SO or O

To the Spigeliaeae sensu Leenhouts (1962) belong the genera *Spigelia*, *Cynoctonum*, *Polypremum*, and *Mitrasacme*. *Cynoctonum* has pollen grains which are characteristic for the Logania type. The species of *Mitrasacme* show different types of pollen grains. *Mitrasacme elata* pollen resemble to the Spigelia-type, *M. indica* and *M. erophila* pollen have several characters in common with the Logania-type, and *M. pygmaea* pollen seem to have some similarity with the Gelsemium-type.

Too few species of *Mitrasacme*—mainly Malesian ones—have been investigated, but as this largely Australian genus is in urgent need of a revision, it seems better to postpone further conclusions. It may be expected that pollen morphology will be of great help in a taxonomic revision.

Illustrations

Erdtman 1943. Pl. IX fig. 148, 149. *Desfontainia spinosa* Ruiz et Pavon.

Erdtman 1952. Fig. 145. D. *Spigelia anthelmia* Linn.

Chromosome numbers (Gadella 1962, Moore 1947)

<i>Desfontainia spinosa</i> Ruiz et Pavon	2n = 14
<i>Polypremum procumbens</i> Linn.	22
<i>Spigelia anthelmia</i> Linn.	32
<i>S. marilandica</i> (Linn.) Linn.	48
<i>S. splendens</i> Wendl. ex Hook.	26

STRYCHNEAE, CYNOCTONUM, LOGANIA AND MITRASACME (p. p.)

LOGANIA-TYPE

Apertures. — Pollen grains 3-colporate, rarely 4-colporate. Colpi sunken, their ends often blunt. Colpus membrane smooth. Ora circular to lalongate (endopori to endocolpi), their ends diffuse. Costae may be present.

Shape. — Equatorial view: grains usually suboblate to oblate-spheroidal, sometimes spheroidal or even prolate-spheroidal (*Cynoctonum* species, *Mitrasacme erophila*). Polar view: grains goniotreme (subangular, semi-angular or sexangular).

Exine. — Sexine mostly thicker than nexine. Tectum smooth, perforate. Some species in *Cynoctonum* have no perforations. Bacules (columellae) low, capita usually distinct.

<i>Cynoctonum mitreola</i>	18 × 18 μ	Sph	perforations in tectum more crowded in the mesocolpia than in the apocolpia
<i>Mitreola</i> (= <i>Cynoctonum</i>) <i>petiolata</i>	15 × 15.5 μ	OSph or Sph	tectum smooth, perforate

<i>Cynoctonum sessilifolia</i>	16 × 14 μ	SP or PSph	bacules indistinct, tectum not perforate
<i>C. sphaerocarpum</i>	15 × 19 μ	SO	colpi narrow, ora distinct, circular; rhombic in equatorial view
<i>Gardneria multiflora</i>			pollen grains too young for measurements
<i>G. nutans</i>	23 × 26 μ	SO or OSph	
<i>G. ovata</i>	21 × 25 μ	SO	
<i>Logania floribunda</i>	20 × 24 μ	SO	costae faint; perforations in tectum rather wide
<i>L. floribunda</i> (syn. <i>L. angustifolia</i>)	25 × 25 μ	Sph	
<i>L. hyssopoides</i>	28 × 29 μ	OSph	no costae; grains semiangular
<i>L. linifolia</i>	23 × 26 μ	SO or OSph	
<i>L. longifolia</i>	25 × 25 μ	Sph	
<i>L. ovata</i> — Pl. 2: 1-4	25 × 25 μ	Sph	
<i>Mitrasacme erophila</i>	32 × 29 μ	PSph	ora indistinct
<i>Neuburgia celebica</i>	32 × 37 μ	SO	costae indistinct
<i>Couthovia</i> (= <i>Neuburgia</i>) <i>collina</i>	26 × 31 μ	SO	
<i>Neuburgia corynocarpa</i>	26 × 31 μ	SO	
<i>Couthovia</i> (= <i>Neuburgia</i>) <i>novocaledonica</i>	25 × 32 μ	SO	
<i>Neuburgia tubiflora</i>	34 × 41 μ	SO	
<i>Strychnos angolensis</i>	23 × 25 μ	OSph	ora circular or slightly longate
<i>S. chrysophila</i>	34 × 39 μ	SO or OSph	
<i>S. mellodora</i>	21 × 23 μ	OSph	
<i>S. nux-vomica</i>	37 × 42 μ	SO or OSph	Ora circular; no costae
<i>S. parviflora</i>	19 × 23 μ	SO	
<i>S. variabilis</i>	36 × 37 μ	OSph	

To this type belong the genera *Logania*, *Cynoctonum*, *Neuburgia*, *Gardneria* and *Strychnos*, and besides, the species *Mitrasacme erophila*.

The *Logania*-type has some characters in common with the *Antonia*-type as well as with the *Spigelia*-type. The differences separating it from the *Antonia*-type are in the ora, from the *Spigelia*-type in the apertures. The type is easily recognisable by its angular shape and smooth tectum. The grains are usually subangular but sometimes semiangular. All other characters are more or less variable. In equatorial view most grains have a polar axis shorter than the equatorial

axis (exceptions in *Cynoctonum*, *Mitrasacme*). The ora are short and hardly or not longer than the colpus width.

The Geniostoma-type and the Spigelia-type have some pollen-morphological characters in common with the Logania-type.

To the tribe Strychnae belong the genera *Strychnos*, *Gardneria*, and *Neuburgia*. In this circumscription the group is accepted by Solereder (1892), Gandoger (1923), Klett (1924), and Leenhouts (1962). Pollenmorphologically related are the genera *Logania*, *Cynoctonum* and some species of *Mitrasacme*, but it seems that in most other characters they are divergent. No taxonomist has suggested this combination of genera before, though Leenhouts (1962) assumes a closer affinity between the Strychnae and the Loganieae via *Neuburgia*.

MITRASACME INDICA-TYPE

Apertures. — Pollen grains 3-colporate. Colpi rather wide and long, sunken, their ends blunt. Colpus membrane smooth. Ora circular, indistinct. No costae present.

Shape. — Equatorial view: grains subprolate. Polar view: grains circular.

Exine. — Sexine thicker than nexine. Tectum smooth, perforate. Bacules (columellae) inordinate to slightly rugulate.

<i>Mitrasacme indica</i>	27 × 25.5 μ	SP	Ora indistinct, no costae
<i>M. neglecta</i>	23 × 25 μ	OSph	Ora circular or slightly longate

The pollen grains of the *Mitrasacme indica*-type show a distinct morphological resemblance to those of *M. erophila*. The pollen of the latter species, however, undoubtedly belong to the Logania-type because of their angular shape, whereas those of the former species are circular in polar view.

MITRASACME PYGMAEA-TYPE

Apertures. — Pollen grains 3-colporate. Colpus ends sharp. Colpus membrane smooth. Ora slightly alongate to circular (endopori), distinct. No costae present.

Shape. — Equatorial view: grains spheroidal or oblate-spheroidal. Polar view: grains circular.

Exine. — Sexine thicker than nexine. Grains reticulate. Lumina fine, smaller than 1 μ. Muri simplibaculate.

<i>Mitrasacme pygmaea</i> var. <i>malaccensis</i>	27.5 × 30 μ	OSph
<i>M. polymorpha</i>	31 × 31 μ	Sph

The pollen grains of *Mitrasacme polymorpha* and *M. pygmaea* resemble those of the *Gelsemium*-type in many characters. *Mitrasacme* grains have, however, a fine reticulum and the longest axis is shorter than in the *Gelsemium*-type.

The genus *Mitrasacme* is in urgent need of a revision. Further investigation of the pollen must be postponed until a new revision is available (see p. 478).

Perhaps the examined specimen of *M. polymorpha* also belongs to *M. pygmaea*, as nearly all specimens of *M. polymorpha* proved to be incorrectly determined.

Illustrations

Wang 1960. Tab. LXII *Gardneria multiflora* Makino, *Strychnos umbellata* (Lour.) Merr.

Chromosome numbers (Gadella 1962, 1963, Janaki-Ammal in Moore 1947) Mohrbutter 1936.

<i>Strychnos aculeata</i> Soler.	$2n = 44$
<i>S. afzelii</i> Gilg	44
<i>S. angolensis</i> Gilg	88
<i>S. camptoneura</i> Gilg	44
<i>S. congolana</i> Gilg	44
<i>S. floribunda</i> Gilg	44
<i>S. icaja</i> Baill.	44
<i>S. laurina</i> DC.	24
<i>S. longicaudata</i> Gilg	44
<i>S. malacoclados</i> C. H. Wright	88
<i>S. nigritana</i> Baker	44
<i>S. nux-vomica</i> Linn.	24
<i>S. soubrensis</i> Hutch. et Dalz.	44
<i>S. spinosa</i> Lam.	44
<i>S. splendens</i> Gilg	44

GELSEMIEAE

GELSEMIUM-TYPE

Apertures. — Pollen grains 3-colporate. Colpi sunken. Colpus membrane smooth in *Mostuea* and with some scattered granules in *Gelsemium*. Ora slightly lalongate (endoporus slightly elongated). The margin of the os is distinctly thickened. Ora not much longer than the colpus width.

Shape. — Equatorial view: grains vary from spheroidal to subprolate. Polar view: grains circular to slightly semiangular, never subangular. Size of the grains is larger than in most other types in the Loganiaceae, usually ca. 40 μ .

Exine. — Sexine many times thicker than nexine. *Mostuea* species are

striate on the tectum. *Gelsemium* species are striato-reticulate, simplibaculate. All *Mostuea* species studied have a tectum perforatum.

<i>Gelsemium elegans</i>	46 × 40 μ	PSph or SP	
<i>G. rankinii</i>	41 × 41 μ	Sph	
— Pl. 2: 8–10			
<i>Mostuea batesii</i>	60 × 51 μ	SP	
<i>M. brunonis</i>	52 × 50 μ	PSph	grains sometimes syncolpate
<i>M. hirsuta</i>	46 × 45 μ	Sph or PSph	
<i>M. surinamensis</i>	46 × 38 μ	SP	
— Pl. 2: 11–12			

Gelsemium and *Mostuea* have pollen grains of this type

The most remarkable characters of the type are the size of the grains, the distinct bacules (columellae), and the well marked margin of the ora. Distinct ora are also found in the Antonia-type, and, less pronouncedly, in the Logania-type. In the Logania-type, however, the ends of the ora are usually diffuse.

The two genera can be distinguished by their difference in exine structure.

- A. Exine striato-reticulate *Gelsemium*
 B. Striae on tectum, tectum
 perforatum, columellae
 inordinate *Mostuea*

Retzia, having indistinct ora, is placed in a type by itself. Several other characters distinguish it also from the *Gelsemium*-type (see p. 481).

Pollen grains of the Oleaceae, which are thought by some authors to be near the *Gelsemieae* are different from those of the *Gelsemium*-type. Although all genera examined had reticulate grains, the reticulation is quite different. The reticulum in *Forestiera*, *Forsythia*, *Schrebera*, and some other genera resembles the reticulum in the *Fagraea ceilanica*-type.

Taxonomic relationship between *Gelsemium* and *Mostuea* is accepted by most taxonomists, viz. Klett (1924), Leeuwenberg (1961), and Leenhouts (1962).

Illustrations

Erdtman 1943. Pl. XIV, fig. 237, *Gelsemium sempervirens* Ait.

Erdtman 1952, fig. 145. C, *Gelsemium sempervirens* Ait.

Wang 1960. Tab. LXII, p. 117, *Gelsemium elegans* (Gardn. et Champ.) Benth.

Chromosome numbers (Moore 1947, Gadella 1962, 1963)

<i>Gelsemium sempervirens</i> Ait.	2n = 16
<i>Mostuea brunonis</i> Dider.	20
<i>M. hirsuta</i> (T. Anders. ex Benth. et Hook.) Baill. ex Baker	20

RETZIEAE

RETZIA-TYPE

Apertures. — Pollen grains 3-colporate. Colpus ends sharp. Colpus membrane densely granulate. Ora indistinct, lalongate (endocolpus). No costae present.

Shape. — Equatorial view: grains oblate-spheroidal to spheroidal. Polar view: grains circular to semiangular.

Exine. — Sexine thicker than nexine. Grains reticulate. Lumina narrower than the muri, muri broad, duplibaculate.

Retzia capensis 43 × 45 μ OSph
— Pl. 3: 1-4

To this type belongs *Retzia* only

The *Retzia*-type resembles the *Gelsemium*-type in several characters. The grains are rather large, 3-colporate, and have a coarse reticulum. There are, however, three important differences: 1. Ora indistinct. 2. Colpus membrane densely granulate. 3. Muri duplibaculate. Especially the indistinct ora separate the *Retzia*-type from the *Gelsemium*-type. Indistinct ora are also characteristic for both types of the *Buddleieae*, but in that tribe granulation of the colpus membrane is never found.

The affinities of *Retzia* are uncertain (Leenhouts, 1962). Leeuwenberg (1964) has the opinion that a relationship to the *Solanaceae* (*Metternichia* and *Sessea*) is less certain than to the *Loganiaceae*. Within the *Loganiaceae* *Retzia* should form a separate tribe near the tribes *Antonieae* and *Buddleieae*. Pollen grains of *Metternichia* and *Sessea* differ widely from *Retzia*; those of the *Antonia* type (*Antonieae*) are also quite different. The pollen grains of the *Buddleieae*, however, have some characters in common with the *Retzia* type.

Neither illustrations nor chromosome numbers published.

LOGANIEAE p. p. (EXCEPT LOGANIA)

GENIOSTOMA-TYPE

Apertures. — Pollen grains porate. Number of pores fluctuates from 3 to 5 and in some rare cases even 6 or 7 pores are present. Pores provided with costae, not protruding, not always in the same plane.

Shape. — Equatorial view: grains oblate to oblate-spheroidal. Polar view: grains distinctly goniotreme (semiangular or subangular).

Geniostoma subtype

Exine. — Sexine thinner than nexine. Tectum smooth, perforate.

<i>Geniostoma balseanum</i>	33 × 37 μ	SO	pollen grains 3-porate, pore diam. ca. 8 μ
<i>G. pancheri</i>	23 × 31 μ	O	pollen grains 3–7 porate, pore diam. ca. 4 μ
<i>G. rupestre</i> (syn. <i>G. australianum</i>) — Pl. 3: 5–7	22 × 28 μ	SO	pollen grains 3–5 porate, pore diam. ca. 5 μ
<i>G. rupestre</i> (syn. <i>G. ligustrifolium</i>)	20 × 25 μ	SO	pollen grains 3-porate, pore diam. 2–3 μ
<i>Labordia hypoleuca</i>	36 × 43 μ	SO	pollen grains 3–5 porate, pore diam. ca. 5 μ. Pores not always in the same plane; sometimes vestibulum present
<i>L. hedyosmifolia</i>	35 × 41 μ	SO	pollen grains 4–5 porate, pore diam. 3–4 μ. Pores not always in the same plane; sometimes a small vestibulum present

Darbolia subtype

Exine. — Sexine thicker than nexine. Grains reticulate. Reticulum very coarse, muri thick, lumina wide.

Labordia helleri According to Selling (1947): pollen grains 3–4-porate (rarely 5-porate), diam. pori ca. 3 μ, reticulum but slightly continuous, lumina about 5 (3–8) μ across.

L. tinifolia var. *tenuifolia* 32 × 40 μ SO pollen grains 3–4-porate, pore diam. ca. 6 μ.
— Pl. 4: 1–5

Geniostoma and *Labordia* have pollen grains of this type.

The *Geniostoma* subtype differs from the *Darbolia* subtype in exine structure only. Superficially the *Geniostoma*-type resembles the *Potalia*-type. Both types are porate, but the *Geniostoma*-type is

distinctly angular, whereas the Potalia-type is more or less spheroidal. The similarity is probably a matter of convergence. The exine structure of the Geniostoma-type and the Potalia-type is similar.

Although the Geniostoma-type and the Darbolia subtype differ considerably in surface pattern, their resemblances in shape and apertures are so striking that it seems better to place both as subtypes in one type.

Taxonomically the genera *Geniostoma* and *Labordia* are distinctly related. Klett (1924) placed *Geniostoma* and *Labordia* in the subtribe Geniostomae. Solereder (1892), Gandoger (1923), and Leenhouts (1962) added *Logania*, forming the tribe Loganieae. Although a certain similarity between the pollen grains of *Logania* and the Geniostoma-type is present, it seems better to place *Logania* next to the representatives of the Strychneae (*Strychnos*, *Gardneria*, and *Neuburgia*).

Illustrations

Erdtman 1948, fig. 6, *Geniostoma rupestre* Forst.

Erdtman 1952, fig. 145 B a-d *Labordia hedyosmifolia* Baill., e *Labordia tinifolia* A. Gray, f-g *Labordia helleri* Sherff.

Selling 1947, Pl. 41, fig. 641-643 *Labordia hedyosmifolia* Baill., 644-645 *Labordia molokaiana*, 646-649 *Labordia waialealae*, 650 *Labordia helleri* Sherff, 651-652 *Labordia tinifolia* A. Gray.

Chromosome numbers (Gadella 1963)

Geniostoma rupestre, Forst. (syn. *G. ligustrifolium*) $2n = 40$

POTALIEAE

POTALIA-TYPE

Apertures. — Pollen grains porate. Number of pores fluctuating between 3 and 5. When more than 3 pores are present the grains are usually stephanoporate. Sometimes the pori are not in the same plane. Pores protruding. Distinct annulus and costae present.

Shape. — Equatorial view: grains spheroidal or oblate spheroidal. Polar view: grains circular.

Exine. — Sexine thicker than nexine. Grains with a smooth usually perforated tectum.

<i>Anthocleista amplexicaulis</i>	34 × 34.5 μ	OSph or Sph	pollen grains 3-(rarely 4-) porate, pore diam. ca. 5 μ
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<i>A. djalonenensis</i>	28 × 28 μ	Sph	pollen grains 3-5-porate, pore diam. ca. 4 μ, pores not always in the same plane. Tectum not perforate
<i>A. grandiflora</i>	30 × 35 μ	SO	pollen grains 3-5-porate, pore diam. ca. 5 μ. Perforations in tectum crowded, transition to reticulum
<i>A. liebrechtsiana</i>	43 × 46 μ	OSph	pollen grains 3-4-porate, pore diam. 6-7 μ. Perforations in tectum crowded, transition to reticulum
<i>A. madagascariensis</i>	33 × 35 μ	OSph	pollen grains 3-4-porate, pore diam. 6-7 μ
<i>A. nobilis</i> — Pl. 4: 9-10	33 × 34 μ	OSph or Sph	pollen grains 3-4 (rarely 5-)porate, pore diam. ca. 4 μ. Perforations in tectum crowded, transition to reticulum
<i>Fagraea gardenioides</i> ssp. <i>borneensis</i>	32 μ (longest axis)		pollen grains 3-4-porate. Tectum not perforate
<i>Potalia amara</i> — Pl. 4: 6-8	39 × 40 μ	OSph or Sph	pollen grains 3-(rarely 4-)porate, pore diam. ca. 4 μ. Tectum not perforate

Potalia and *Anthocleista* and the species *Fagraea gardenioides* ssp. *borneensis* have pollen grains of this type

The *Potalia*-type shows a superficial resemblance to the *Geniostoma* type. Both types are porate, have a smooth tectum, and may have different numbers of pores. This similarity, however, is in my opinion only a convergence. In the *Potalia*-type the grains are spheroidal or oval in shape, in the *Geniostoma*-type angular.

It is uncertain which type of the *Loganiaceae* is morphologically closest to the *Potalia*-type. The exine structure and the pori of the *Antonia*-type have some similarity. If we consider reduction of colpi as a morphological evolution in *Loganiaceae* (see *Fagraea*), the *Antonia*-type with its circular ora (endopori) is probably closest to the *Potalia*-type.

Pollen grains of *Potalia* and *Anthocleista* are very much alike. The pori in *Potalia amara* are somewhat more protuberant than in the *Anthocleista* species, but this is a gradual difference only. Pollen

grains of *Fagraea gardenioides* ssp. *borneensis* cannot be separated from *Anthocleista* pollen. They seem to be a perfect link between the *Potalia*- and the *Fagraea*-types.

Taxonomists regard *Potalia* and *Anthocleista* as distinctly related (Solereeder 1892; Gandoger 1923; Klett 1924; Leeuwenberg 1961; Leenhouts 1962). They also suggest a close relationship with *Fagraea*. As pollen grains of *Fagraea gardenioides* ssp. *borneensis* belong to the *Potalia*-type and, on the other hand, are distinctly related to some pollen types in *Fagraea* (*F. berteriana*-type), this relationship seems justified.

Closer affinity of the Loganiaceae-Potalieae with the Apocynaceae-Tabernaemontanae has been suggested (Leenhouts, 1962); the pollen grains are, however, distinctly different.

Illustrations

Erdtman 1952, fig. 145 A. *Anthocleista parviflora* ("parvifolia") (syn. = *A. nobilis* G. Don; Leeuwenberg, 1961).

Chromosome numbers (Gadella 1961, 1963)

<i>Anthocleista djalonensis</i> Cheval.	2n = 60
<i>A. liebrechtsiana</i> Wildem. et Durand	60

FAGRAEA FRAGRANS-TYPE

Fagraea fragrans subtype

Apertures. — Pollen grains 3-colporate. Colpi narrow, rather short. Ora circular (endopori). Costae present.

Shape. — Equatorial view: grains spheroidal to oblate-spheroidal. Polar view: grains circular.

Exine. — Sexine thicker than nexine. Grains reticulate, slightly striato-reticulate over the entire surface. Lumina small, not exceeding $2\ \mu$ in width, muri sharp-edged in the upper part, simplibaculate.

Fagraea fragrans $17.5 \times 18\ \mu$ OSph
— Pl. 6: 5-6

Fagraea crenulata subtype

Apertures. — Pollen grains 3-colporate, nearly porate. Colpi very short and indistinct. Ora circular (endopori). Costae present.

Shape. — Equatorial view: grains oblate-spheroidal. Polar view: grains circular.

<i>F. longiflora</i>	34 × 37 μ	OSph	pore diam. ca. 4 μ
<i>F. macroscypha</i>	40 × 42 μ	OSph	pore diam. 2–3 μ, capita slightly rounded at the top
<i>F. resinosa</i>	38 × 43 μ	O or OSph	pore diam. 5–6 μ
<i>F. tacapala</i> var. <i>gracilis</i>	34 × 38.5 μ	SO or OSph	pore diam. ca. 5 μ
<i>F. tubulosa</i>	47 × 51 μ	OSph	pore diam. 5–6 μ.
<i>F. woodiana</i>	30.5 × 32.5 μ	OSph	pore diam. ca. 4 μ

Fagraea annulata subtype

Apertures. — Pollen grains 3-porate. Pori circular to oval. Costae pori present. Pori wider than one lumen of the reticulum (ca. 2–5 ×).

Shape. — Equatorial view: grains oblate-spheroidal. Polar view: grains circular.

Exine. — Sexine thicker than nexine. Grains coarsely reticulate. Lumina wide, can be ca. 6 μ across, muri simplibaculate or sometimes duplibaculate, lower part (columellae) slender, upper part more or less rounded.

<i>Fagraea annulata</i> — Pl. 5: 1	48 × 48 μ	Sph	pore diam. ca. 8 μ
<i>F. carlsensis</i> — Pl. 5: 2	41 × 44 μ	OSph	pore diam. 6–8 μ, elliptic
<i>F. eymae</i> — Pl. 6: 4	43 × 46 μ	OSph	pore diam. 5–7 μ

FAGRAEA BERTERIANA-TYPE

Apertures. — Pollen grains 3-(rarely 4-)porate or colporate. If colporate colpi very short, their ends a little blunt. Ora circular (endopori). Costae present.

Shape. — Equatorial view: grains suboblate to oblate-spheroidal. Polar view: grains circular.

Exine. — Sexine thicker than nexine. Grains finely reticulate. Lumina small (less than 1 μ) and irregular, but all of about the same size, muri simplibaculate, upper part rounded.

<i>Fagraea berteriana</i>	30 × 34 μ	SO or OSph	pore diam. 3–4 μ
<i>F. berteriana</i> (syn. <i>F. schlechteri</i>) — Pl. 5: 3–4	28 × 31 μ	SO	pollen grains colporate, colpi short
<i>F. bodenii</i>	39.5 × 42.5 μ	OSph	
<i>F. carnosa</i>	40 × 42 μ	OSph	pore diam. 5–6 μ
<i>F. salticola</i>	37 × 42 μ	OSph	pollen grains colporate, pore diam. 5–6 μ, colpi short
— Pl. 6: 10–11			

The types and subtypes in the genus *Fagraea* can be distinguished by their exine patterns. All *Fagraea* species except one (*F. gardenioides* ssp. *borneensis*) have a reticulate exine (the reticulum varies in the different types and subtypes). Points of similarity are found

in the shape and the apertures. The colporate subtypes have circular ones. The porate ones are probably derived from the colporate subtypes by reduction of the colpi. In favour of this theory is the presence of colporate and porate grains, respectively, in two specimens of *F. berteriana*. Besides, the colpi in the colporate specimen are very short.

The *F. fragrans* subtype, *F. crenulata* subtype and *F. racemosa* subtype are related in their reticulum. In the *F. fragrans* subtype and *F. crenulata* subtype a striato-reticulate pattern is present in the apocolpia as well as in the mesocolpia. A distinct striato-reticulate pattern occurs in the *F. racemosa* subtype in the apocolpia only. The *F. crenulata* subtype differs from the *F. fragrans* subtype by very short, indistinct colpi. All subtypes have sharp-edged muri, a character also occurring in the *F. ceilanica* subtype.

The *F. ceilanica* subtype and the *F. annulata* subtype are placed together in one type because both have porate grains and a comparatively coarse reticulum. The typical sharp-edged muri, occurring in the *F. ceilanica* subtype and also present in the *F. fragrans* subtype, are, however, missing in the *F. annulata* subtype, where the muri are rounded in the upper part.

In the *F. annulata* subtype *F. carstensensis* represents a transition to the *F. ceilanica* subtype and *F. eymae* a transition to the *F. berteriana*-type. In *F. carstensensis* the columellae are thicker than in *F. annulata*, but thinner than in the *F. ceilanica* subtype. *F. eymae* grains have a reticulum with narrower lumina than in *F. annulata* but wider than in the *F. berteriana*-type.

The *F. berteriana*-type can be distinguished by its fine reticulum. As in the *F. annulata* subtype the upper part of the muri is rounded. In one specimen of *F. berteriana* the grains are not porate but colporate. The colpi, however, are very short. In *F. bodenii* sometimes pollen grains with four pores occur.

In pollen morphology there is a close similarity between *F. berteriana* and *F. gardenioides* ssp. *borneensis*. The pollen grains of the latter species are not reticulate, however, and belong to the Potalia-type (see page 485).

It is striking that in *Fagraea* the species show such a large variation in their pollen characters. This is in contrast with the relative uniformness of most genera in the Loganiaceae (another example is *Mitrasacme*).

In comparing the different types and subtypes with each other,

it is possible to distinguish several "evolutionary" trends: 1. Reduction of the colpi. 2. Reduction of the lumina of the reticulum. 3. Enlargement of the lumina of the reticulum.

If tricolporate reticulate pollen grains are regarded as more primitive than porate tectate ones, it follows that the *F. fragrans* subtype is the most primitive one in *Fagraea*. This is in accordance with the taxonomic results of Leenhouts (1962). With this subtype as starting point it is possible to draft the following morphological series:

- | | | |
|---|---|--|
| A. <i>F. fragrans</i> → <i>F. crenulata</i> | { <i>F. ceilanica</i>
<i>F. annulata</i> | { Reduction of the colpi
and enlarging of the
lumina |
| B. <i>F. fragrans</i> → <i>F. berteriana</i> → <i>F. gardenioides</i> | | { Reduction of the colpi
and lumina |
| C. <i>F. fragrans</i> → <i>F. racemosa</i> | | Reduction of the lumina |

Most *Fagraea* types are completely different from other Loganiaceae types with regard to exine structures. Shape and apertures are similar in the Potalia-type and Antonia-type and to those occurring in the porate pollen types of *Fagraea*.

Pollen grains in the Apocynaceae-Tabernaemontanae (*Tabernaemontana*, *Voacanga*, and *Hazunta*) are different from those in the Potalieae. This result does not support the suggestion that the Potalieae may be taxonomically related to that tribe of the Apocynaceae. On the other hand, some genera in the Oleaceae (viz. *Forestiera*, *Forsythia*, *Schrebera*, and some other genera) have an exine structure very similar to that met within the *F. ceilanica*-type. The apertures in the Oleaceous genera are, however, quite different.

Professor Rowley of the Department of Botany, University of Massachusetts, Amherst, kindly made some electron micrographs of *Fagraea blumei* and *Fagraea ceilanica*. Both species seemed to be identical in their morphological characters when examined with an ordinary light microscope. The replicas distinctly show that the muri of the reticula are different. Both species show a sharp-edged upper part of the muri, but the muri sides in *F. ceilanica* are smooth whereas those in *F. blumei* have small excrescences. Perhaps this character will prove to be of value in a further subdivision of the *F. ceilanica* type, but the making of replicas required much material and is very time-consuming. For these reasons a further investiga-

tion of the *Fagraea* species with the aid of an electron-microscope was not possible.

Illustrations

Wang 1960, pag. 117, tab. LXII, *Fagraea chinensis* (syn. *F. ceilanica* Thunb., Leenhouts, 1962).

Chromosome numbers (Gadella 1963, Mohrbutter 1963)

<i>Fagraea ceilanica</i> Thunb.	$2n = 66$
<i>F. fragrans</i> Roxb.	12
<i>F. littoralis</i> (= <i>F. ceilanica</i> Thunb.)	12

PLOCOSPERMA (Gen. inc. sed.)

PLOCOSPERMA-TYPE

Apertures. — Pollen grains 3-colporate. Colpi long.

Shape. — Equatorial view: grains probably subprolate or prolate; polar axis longer than equatorial axis. Polar view: grains circular.

Exine. — Sexine much thicker than nexine. Pollen grains reticulate. Lumina small, ca. 1μ , simplibaculate, bacules high.

Plocosperma microphyllum: pollen grains immature

Morphologically the pollen grains of *Plocosperma* do not seem related to any pollen type in the Loganiaceae. The apertures differ greatly and the exine structure only shows similarity to that of the grains in the Gelsemium- and Retzia-types. These types, however, differ in too many other characters to suppose a pollen morphological relationship.

Key to pollen types

- 1 a Pollen grains reticulate, striate or striato-reticulate 2
- b Pollen grains smooth 15
- 2 a Pollen grains porate or colpate 3
- b Pollen grains colporate 7
- 3 a Reticulum coarse (lumina $> 3 \mu$) 4
- b Reticulum fine (lumina $< 3 \mu$) 6
- 4 a Pollen grains angular in polar view;
 usually more than three pores Darbolia subtype
- b Pollen grains circular in polar view, triporate 5

- 5 a Upper part of muri sharp-edged . *Fagraea ceilanica* subtype
- b Upper part of muri rounded . . . *Fagraea annulata* subtype
- 6 a Pollen grains goniotreme with
 - apertures at the angles *Mitrasacme elata*-type
 - b Pollen grains circular in polar view *Fagraea berteriana*-type
- 7 a Colpi very short 8
- b Length of colpi at least 3 × diameter of ora 9
- 8 a Reticulum fine (lumina < 2 μ)
 - Upper part of muri rounded . . . *Fagraea berteriana*-type
 - b Reticulum coarse (lumina > 2 μ)
 - Upper part of muri sharp-edged . . . *Fagraea crenulata* subtype
- 9 a Apocolpia striate. *Mesocolpia*
 - reticulate *Fagraea racemosa* subtype
 - b Pollen grains not striate or if striate,
 - striation not in apocolpia only 10
- 10 a Colpus membrane densely granulate.
 - Ora indistinct *Retzia*-type
 - b Colpus membrane not granulate or
 - rarely with a few scattered granules (*Gelsemium*-type) . . . 11
- 11 a Pollen grains with indistinct ora 12
- b Pollen grains with distinct ora 13
- 12 a Pollen grains goniotreme with
 - apertures at the angles in polar view.
 - Ora lolongate *Mitrasacme elata*-type
 - b Pollen grains circular in polar view.
 - Ora lalongate *Chilianthus* subtype
- 13 a Ora circular. Grains small,
 - striato-reticulate *Fagraea fragrans* subtype
 - b Ora slightly lalongate 14
- 14 a Pollen grains finely reticulate
 - (lumina < 1 μ) *Mitrasacme pygmaea*-type
 - b Pollen grains striate or striato-reticulate.
 - Longest axis > 25 μ *Gelsemium*-type
- 15 a Pollen grains porate 16
- b Pollen grains colporate or colpate 17
- 16 a Pollen grains angular in polar view . . . *Geniostoma* subtype
- b Pollen grains circular in polar view.
 - Pores protruding *Potalia*-type
- 17 a Pollen grains colpate or, if colporate,
 - ora lolongate *Spigelia*-type

- b Pollen grains colporate. Ora circular or lalongate 18
 18 a Pollen grains angular in polar view Logania-type
 b Pollen grains circular in polar view 19
 19 a Ora distinct, circular or slightly lalongate Antonia-type
 b Ora indistinct, circular or lalongate 20
 20 a Ora circular Mitrasacme indica-type
 b Ora lalongate 21
 21 a Pollen grains small (longest axis not exceeding
 25 μ) 3- or 4- colporate. Bacula short,
 sometimes indistinct. Buddleia subtype
 b Pollen grains larger, 4-colporate.
 Bacula distinct Emorya-type

Table 1. Specimens investigated

Ma = Macromethod. Mi = Micromethod. NPC = number, position, character of apertures

Name	Method	NPC
Antonieae		
<i>Antonia ovata</i> Pohl	B. W. 4355; U	Ma 345
<i>Bonyunia antoniiifolia</i> Progel	Ducke 12197; U	Mi 345
<i>B. aquatica</i> Ducke	Ducke 354; A	Mi 345
<i>B. minor</i> N. E. Brown	Lasser 1472; VEN	Mi 345
<i>B. superba</i> R. Schomb.	Schomburgk 614; P	Mi 345
<i>Norrisia maior</i> Soler.	Herb. Sandakan 25274; L	Mi 345
<i>N. malaccensis</i> Gardner	Griffith 3731; S	Ma 345
<i>Usteria guineensis</i> Willd.	Voorhoeve 106; WAG	Mi 345
Buddleieae		
<i>Adenoplea sinuata</i> Radlk.	D'Alleizette s.n.; L	Mi 345
	Afzelius s.n.; SPL	Ma 345
<i>Adenoplusia axillaris</i> Radlk.	Hildebrandt 3671; SPL	Ma 345
<i>A. uluguruensis</i> Melch.	Schlieben 2756 (isotype); P	Mi 345
<i>Androya decaryi</i> Perrier	Serv. Forest. 8488; P	Mi 343/345
<i>Buddleia alpina</i> Oerst.	P. H. Allen 3458; U	Ma 345/445
<i>B. americana</i> Linn.	Rutten & Rutten-Pekelhar- ing 134; U	Ma 345/445
<i>B. coriacea</i> Remy	Brooke 6382; U	Ma 345
<i>B. corrugata</i> (Benth.) Phill.	Drège s.n. (3618?), anno 1837; S	Ma 345
<i>B. dysophylla</i> (Benth.) Radlk.	Verdcourt s.n., anno 1954; K (SPL)	Ma 345

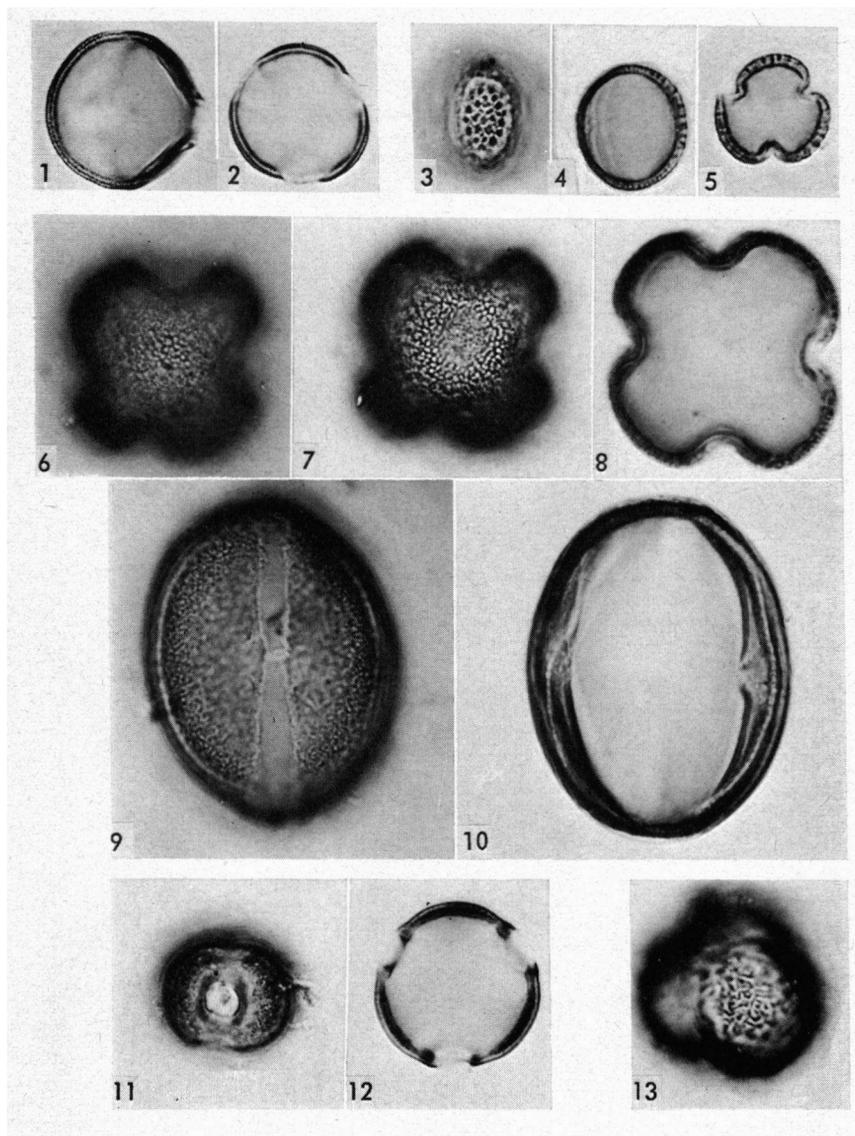
Name	Method	NPC
<i>B. elegans</i> Cham. et Schlecht.	Hatschbach 8298; U	Ma 345
<i>B. glomerata</i> Wendl. f.	Coll. ?, S. Africa, anno 1824; S	Ma 345
<i>B. indica</i> Lam.	Leeuwenberg 3511; WAG	Mi 345
<i>B. madagascariensis</i> Lam.	De Cary 10061; L	Mi 345
<i>B. marubiiifolia</i> Benth.	Hinckley 3098; U	Ma 345
<i>B. saligna</i> Willd.	Scott Elliot 356; U	Mi 345
<i>B. salviifolia</i> (Linn.) Lam.	Stolz 2068; U	Mi 345
<i>B. stachyoides</i> Cham. et Schlecht.	R. Klein 560; U	Ma 345
<i>Emorya suaveolens</i> Torrey	Purpus 4748; BM	Mi 345/445
<i>Gomphostigma virgatum</i> (Linn.) Kuntze	Pont 1718; U	Ma 345
—	Norlindh & Weimarck 4499; LD	Mi 345
<i>Nuxia capitata</i> Baker	Bahon 3650 (isotype); P	Mi 345
<i>N. congesta</i> R. Brown ex Fresen	Breteler 2573; WAG	Ma 345
<i>N. floribunda</i> Benth. [syn. <i>N. polyantha</i>]	Stolz 2068; U	Ma 345
<i>N. keniensis</i> T. C. E. Fries [syn. <i>N. congesta</i>]	Fries & Fries 777 (type); UPS	Ma 345
<i>N. oppositifolia</i> (Hochst.) Benth. [syn. <i>N. antunesii</i>]	Baum 29 (type); S	Ma 345
<i>N. verticillata</i> Lam.	Johnston s.n., anno 1889; U	Mi 345
<i>Pellanthera floribunda</i> Benth.	A. E. Lawrence 432; A	Mi 345
<i>Sanango durum</i> Bunting et Duke	Wurdack 2018; US	Mi 345
Spigeliaceae (p.p.)		
<i>Desfontainia spinosa</i> Ruiz et Pavon	Leeuwenberg 3507; WAG	Mi 345 [344]
<i>Mitrasacme elata</i> R. Brown	Brass 27227; L	Mi 343
<i>Polypremum procumbens</i> Linn.	Kramer & Hekking 2547; U	Mi 345 [343]
<i>Spigelia anthelmia</i> Linn.	Jonker-Verhoef & Jonker 17; U	Mi 343
<i>S. leiocarpa</i> Benth.	Y. Mexia 6290; U	Mi 343
<i>S. martiana</i> Cham.	Hatschbach 8458; U	Mi 343
Strychnaceae, Loganiaceae (p.p.), Mitrasacme (p.p.)		
<i>Cynoctonum mitreola</i> (Linn.) Britt.	Smitinand 3565; L	Mi 345
<i>Mitreola</i> (= <i>Cynoctonum</i>) <i>petiolata</i> Torrey	E. Wall/Fischer s.n., anno 1933; S	Ma 345

Name		Method	NPC
<i>Cynoctonum sessilifolia</i> J. F. Gmelin	Vesterland s.n., anno 1889; S	Ma	345
<i>C. sphaerocarpum</i> Leenh.	J. & M. S. Clemens 29649 (type); L	Mi	345
<i>Gardneria multiflora</i> Makino	Ren Chang Ching 2955; S	Ma	345
<i>G. nutans</i> Sieb. et Zucc.	Coll. ? 459; L	Ma	345
<i>G. ovata</i> Wall.	Hohenacker 1445; U	Ma	345
<i>Logania angustifolia</i> R. Brown	Watson 23; U	Ma	345
<i>L. floribunda</i> R. Brown	Constable 7242; U	Ma	345
<i>L. hyssopoides</i> Nees	Mueller s.n.; U	Mi	345
<i>L. tinifolia</i> Schlecht.	Kaspiew 6; U	Ma	345
<i>L. longifolia</i> R. Brown	Coll. ? S. Austr.; U	Ma	345
<i>L. ovata</i> R. Brown	Cunningham 252; U	Ma	345
<i>Mitrasacme erophila</i> Leenh.	Hook. f. & Thomson s.n., Mt. Khasia; L	Mi	345
<i>M. indica</i> Wight	Hook. f. & Thomson s.n., Malabar; L	Mi	345
<i>M. neglecta</i> Leenh.	Coert 917; L	Mi	345
<i>M. polymorpha</i> R. Brown	Constable 6239; U	Mi	345
<i>M. pygmaea</i> R. Brown var. <i>malaccensis</i> (Wight) Hara	Bünnemeyer 5770; U	Mi	345
<i>Neuburgia celebica</i> (Koord.) Leenh.	Kjellberg 2025; SPL	Ma	345
<i>Couthovia</i> (= <i>Neuburgia</i>) <i>collina</i> A. C. Smith	A. C. Smith 6157; S	Ma	345
<i>Neuburgia corynocarpa</i> (A. Gray) Leenh.	A. C. Smith 6330; S	Ma	345
<i>Couthovia</i> (= <i>Neuburgia</i>) <i>novo-caledonica</i> Gilg et Bened.	Däniker 1068; SPL	Ma	345
<i>Neuburgia tubiflora</i> Blume	Aet 143 (type); L	Mi	345
<i>Strychnos angolensis</i> Gilg	Welwitsch 4776 (type); LISU	Ma	345
<i>S. chrysophylla</i> Gilg	Le Testu 9377; P	Ma	345
<i>S. mellodora</i> S. Moore	Hack 4; FHO	Ma	345
<i>S. nux-vomica</i> Linn.	Cult. Hort. Bog. I. C. 6; U	Mi	345
<i>S. parviflora</i> Spruce ex Benth.	Ducke 1110; IAN	Ma	345
<i>S. variabilis</i> De Wildeman	A. Carlier 57; BR	Ma	345
Gelsemieae			
<i>Gelsemium elegans</i> (Gardn. et Champ.) Benth.	Balansa 1041; L	Ma	345
<i>G. rankinii</i> Small	Godfrey 56366; GB	Mi	345
<i>Mostuea batesii</i> Baker	Breteler 2750; WAG	Mi	345

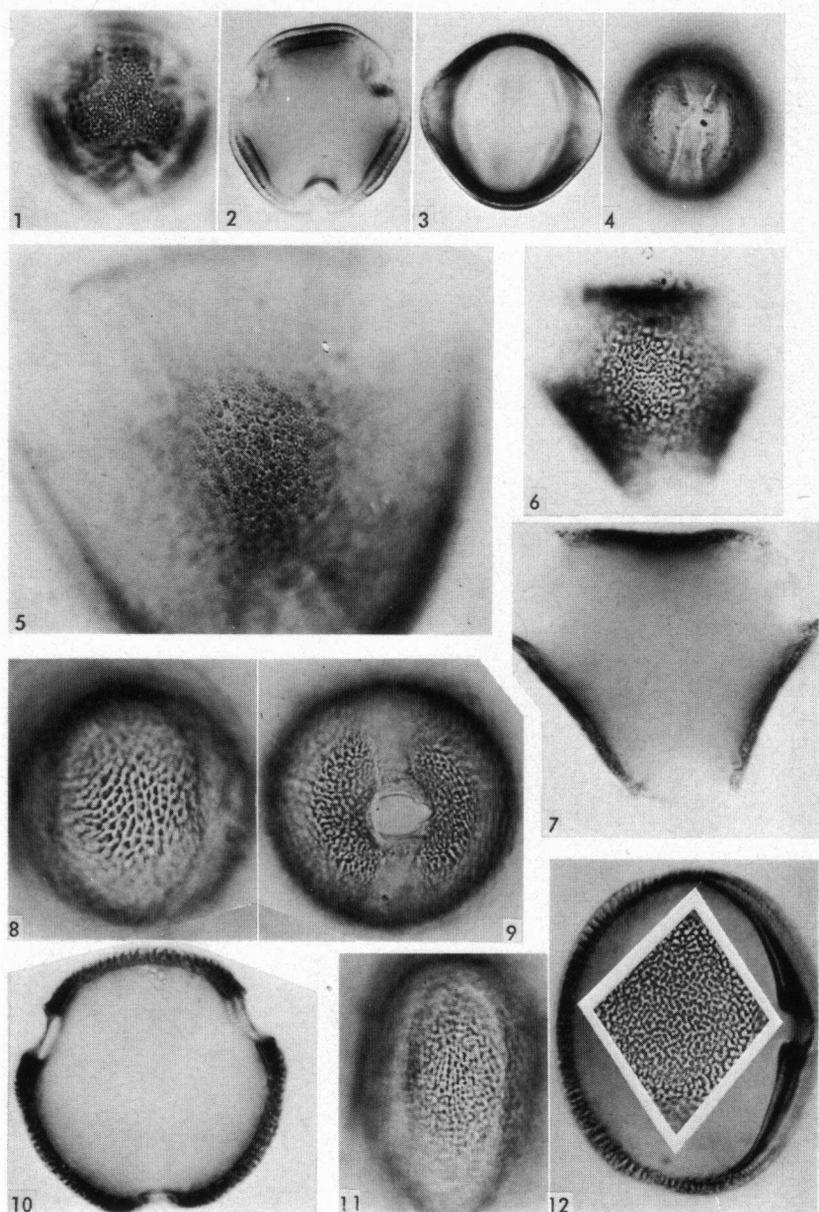
Name		Method	NPC
<i>M. brunonis</i> Dider.	Breteler 2331; WAG	Mi	245/345
<i>M. hirsuta</i> (T. Anders. ex Benth. et Hook.) Baill. ex Baker	Thomas 265; K (SPL)	Ma	345
<i>M. surinamensis</i> Benth.	Versteeg 878; U	Mi	345
Retzieae			
<i>Retzia capensis</i> Thunb.	Bos 674; WAG	Mi	345
Loganieae (p.p.)			
<i>Geniostoma balseanum</i> Baill.	Mackee 5429; L	Mi	344
<i>G. pancheri</i> Baill.	Däniker 3094; SPL	Ma	344/444-744
<i>G. rupestre</i> Forster (syn. <i>G. australianum</i>)	Mueller s.n. (Rockhampton Bay); U	Ma	344/544-744
<i>G. rupestre</i> Forster (syn. <i>G. ligustrifolium</i>)	H. Powell s.n., 2.IX.1947; U	Ma	344
<i>Labordia hedyosmifolia</i> Baill.	Selling s.n., Hawaii, 10.IX.1938; SPL	Ma	444/544
<i>L. hypoleuca</i> Degener	Degener 10,000; SPL	Ma	444/544
<i>L. helleri</i> Sherff	A. A. Heller 2579; BISH	Ma	344/444-644
<i>L. tinifolia</i> A. Gray var. <i>tenuifolia</i> Degener et Sherff ex Sherff	Degener 10272; SPL	Ma	344/444
Potalieae			
<i>Anthocleista amplexicaulis</i> Cheval.	Serv. Forest. 7388; P	Ma	344/444
<i>A. djalonensis</i> Cheval.	Leeuwenberg 3316; WAG	Mi	344/444-544
<i>A. grandiflora</i> Gilg	Stolz 497; S	Ma	344/444-544
<i>A. liebrechtsiana</i> De Wilde-man et Dur.	Breteler 2806; WAG	Mi	344/444
<i>A. madagascariensis</i> Baker	Humbert 23018; P	Ma	344/444
<i>A. nobilis</i> G. Don	Leeuwenberg 2322; WAG	Mi	344/444
<i>Fagraea acuminatissima</i> Merr.	Jacobs 5633; L	Mi	344
<i>F. annulata</i> Hiern	Versteeg 1237 (type); U	Mi	344
<i>F. auriculata</i> Jack ssp. <i>borneensis</i> (Sheff.) Leenh.	Cult. Hort. Bog.; U	Mi	345
<i>F. berteriana</i> A. Gray ex Benth.	Brass 28140; L	Ma	344
<i>F. berteriana</i> A. Gray ex Benth. (syn. <i>F. Schlechteri</i>)	Däniker 648; SPL	Ma	345
<i>F. blumei</i> G. Don	Koorders 4321 b; S	Ma	344
<i>F. bodenii</i> Wernh.	Pulle 894; U	Mi	345
<i>F. carnosa</i> Jack	Cult. Hort. Bog. X.G. 24; L	Mi	344

Name		Method	NPC
<i>F. carstensensis</i> Wernh.	P. v. Royen & Sleumer 7755; L	Mi	344
<i>F. ceilanica</i> Thunb.	Hulstijn 363; L	Mi	344
<i>F. crenulata</i> Clark	Hort. Bot. Singapore 1904; L	Mi	345
<i>F. elliptica</i> Roxb.	Hort. Bog. 592; L	Ma	345
<i>F. eymae</i> Backer	Eyma 4330 bis (type); L	Ma	344
<i>F. fastigiata</i> Blume	Coll. ? Herb. No 908.127-108; L	Ma	344
<i>F. fragrans</i> Roxb.	Kostermans 8955; L	Ma	345
<i>F. gardenioides</i> Ridl. ssp. <i>borneensis</i> Leenh.	Sarawak Museum 2508 (type); L	Mi	344
<i>F. gracilipes</i> A. Gray	Robins 570; L	Mi	345
<i>F. involucrata</i> Merr.	Sandakan Herb. 16380; L	Ma	344
<i>F. aff. longiflora</i> Merr.	Elmer 9159; L	Ma	344
<i>F. macroscypha</i> Baker	?; L	Ma	344
<i>F. racemosa</i> Jack ex Wall.	Santos 4256; L	Ma	345
<i>F. resinosa</i> Leenh.	Hallier 3162 (type); L	Ma	344
<i>F. salticola</i> Leenh.	Hoogland & Schodde 7425; L	Mi	344
<i>F. tacapala</i> Leenh. ssp. <i>gracilis</i> Leenh.	Kjellberg 1521 a (type); S	Ma	344
<i>F. tubulosa</i> Blume	Coll. ? (type); L	Mi	344
<i>F. umbelliflora</i> Gilg et Bened.	Ledermann 9614 (type); L	Mi	345
<i>F. woodiana</i> F. v. Muell.	J. J. F. E. de Wilde 1202; L	Mi	344
<i>Potalia amara</i> Aubl.	Versteeg 323; U	Mi	344/444
<i>Plocosperma</i> (gen. inc.sed.)			
<i>Plocosperma microphyllum</i> Baill. ex Soler.	Galeotti (type); P [SPL]	Ma	343

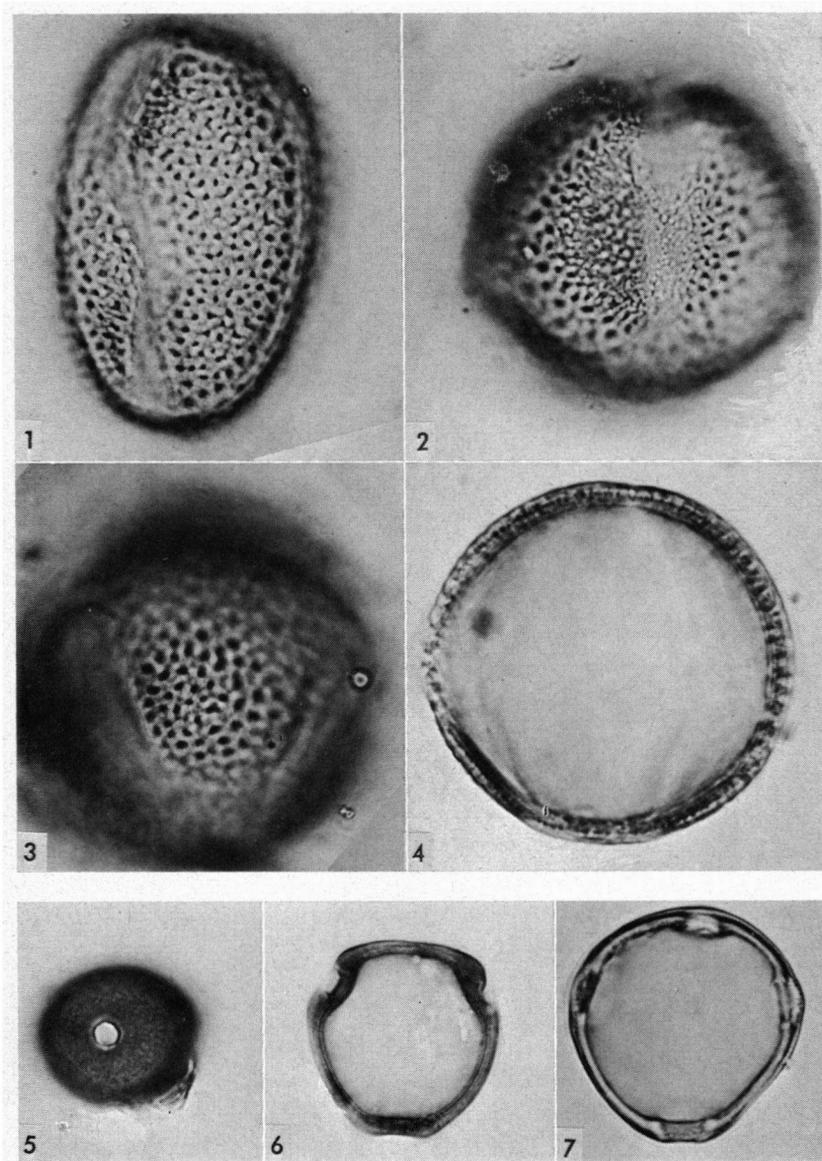
Botanisch Museum en Herbarium, Utrecht, Netherlands.



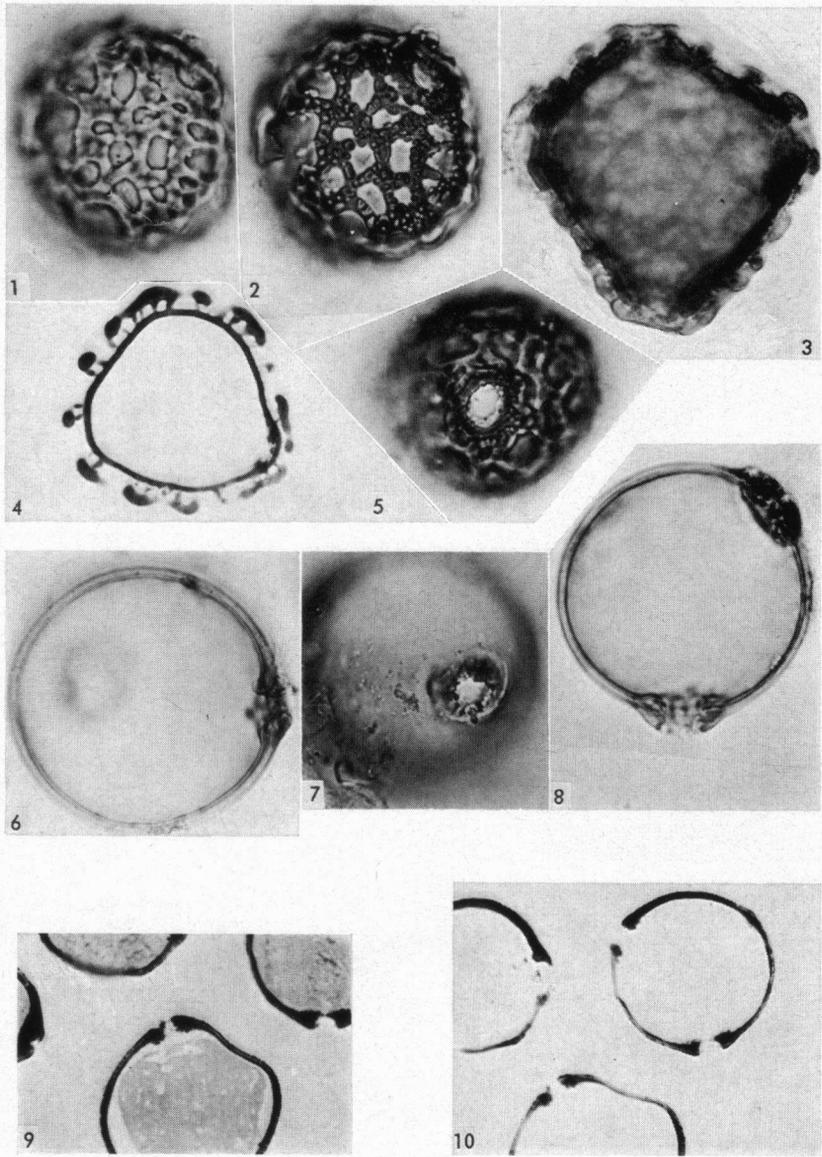
PL. 1. 1-2, *Buddleia*-type, *Buddleia* subtype (*Buddleia indica*).—3-5, *Buddleia*-type, *Chilanthus* subtype (*Buddleia saligna*).—6-10, *Emorya*-type (*Emorya suaveolens*).—11-13, *Antonia*-type. 11-12 (*Antonia ovata*). 13 (*Bonyunia aqualica*). $\times 1200$.



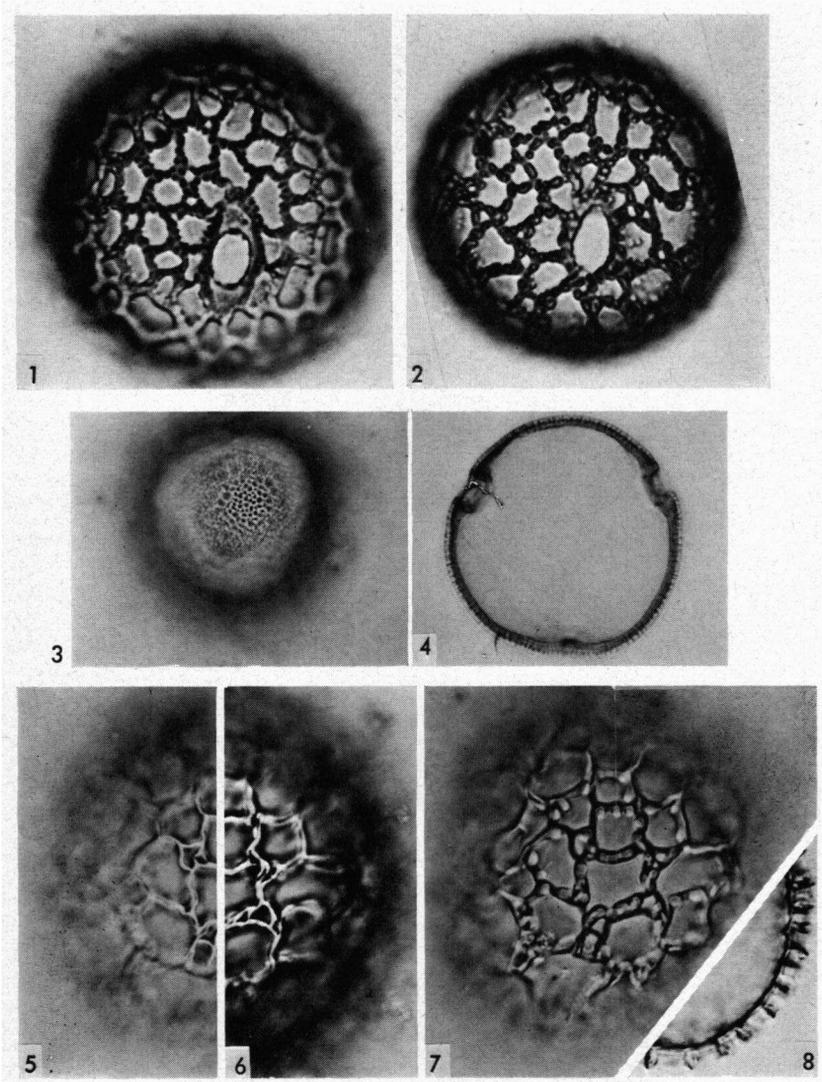
Pl. 2. 1-4, *Logania*-type (*Logania ovata*).—5-7, *Spigelia*-type. 5 (*Spigelia leiocarpa*). 6-7 (*Desfontainia spinosa*).—8-12 *Gelsemium*-type. 8-10 (*Gelsemium rankinii*). 11-12 *Mostuea surinamensis*. $\times 1000$.



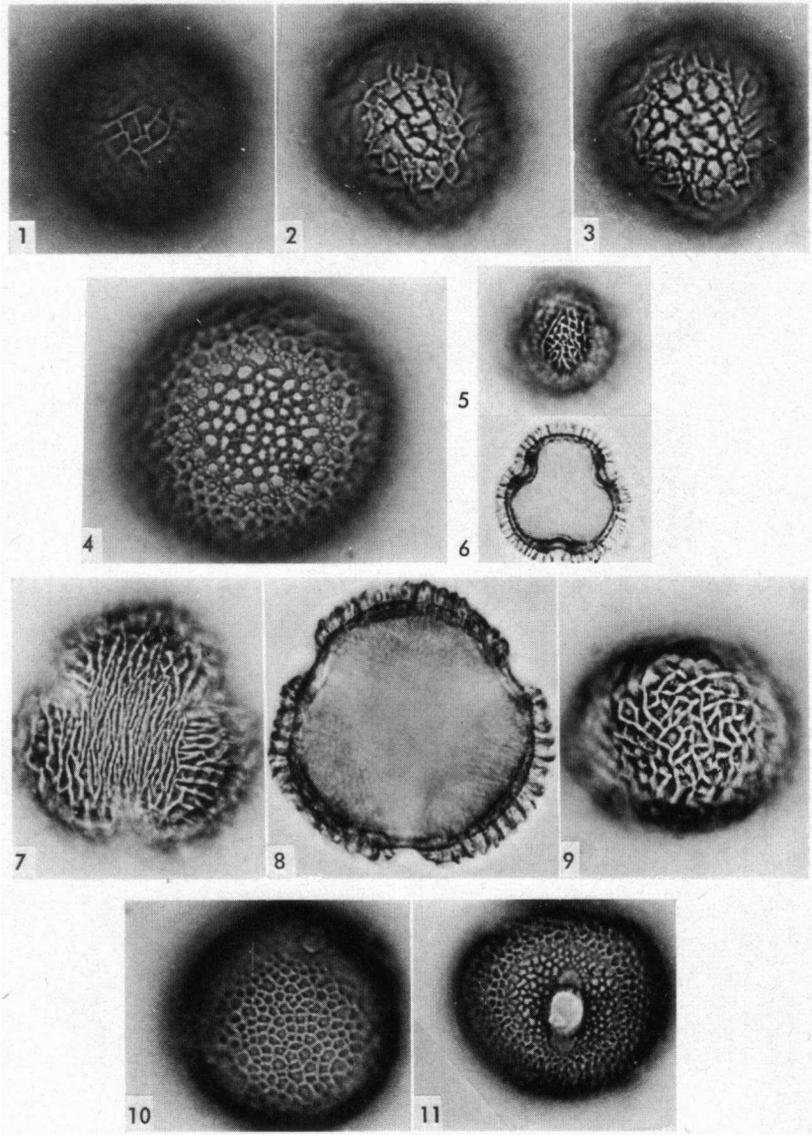
PL. 3. 1-4, *Retzia*-type (*Retzia capensis*).—5-7, *Geniostoma*-type, *Geniostoma* subtype (*Geniostoma rupestre*). $\times 1200$.



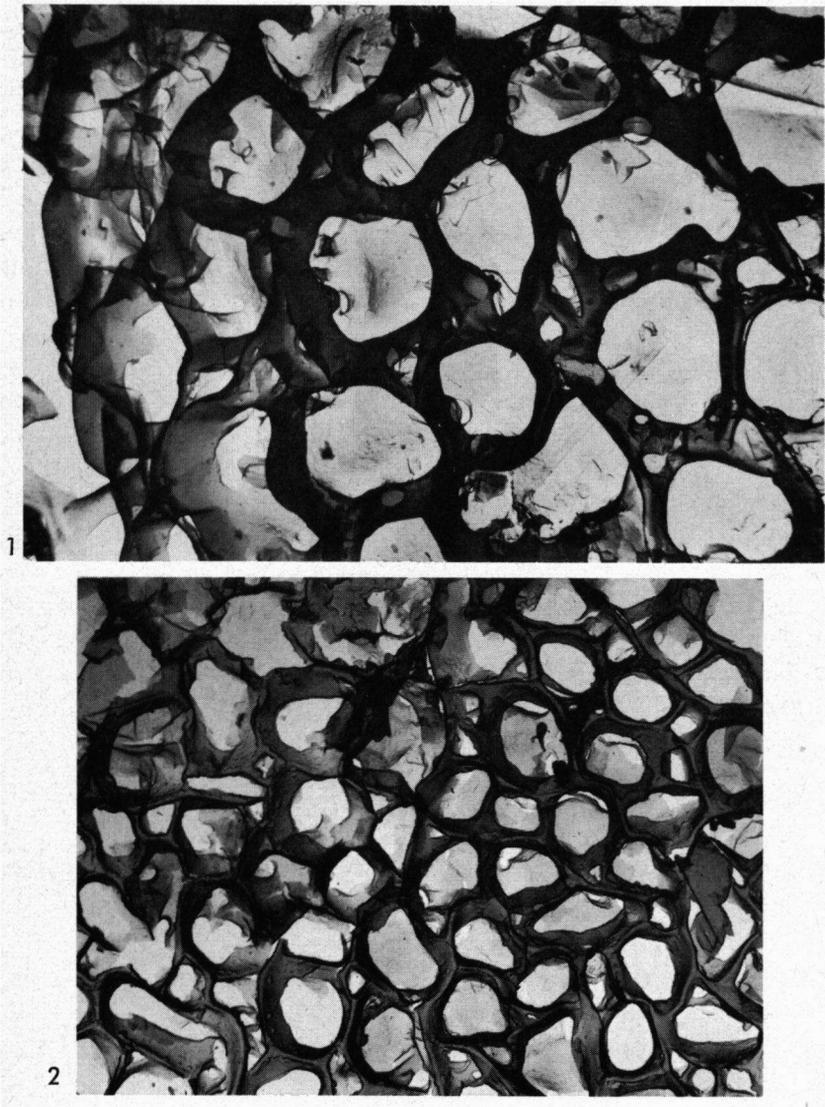
Pl. 4. 1-5, Geniostoma-type, Darbolia subtype (*Labordia tinifolia*).—6-10, Potalia-type. 6-8 (*Potalia amara*), 9-10 (*Anthocleista nobilis*), section about 0.5μ thick. $\times 1000$.



Pl. 5. 1-2, *Fagraea ceilanica*-type, *Fagraea annulata* subtype. 1, (*F. annulata*). 2, (*F. carstensensis*).—3, *Fagraea berteriana*-type (*F. berteriana*).—5-8, *Fagraea ceilanica*-type, *Fagraea ceilanica* subtype (*F. ceilanica*). $\times 1100$.



Pl. 6. 1-3, *Fagraea fragrans*-type, *Fagraea crenulata* subtype (*F. crenulata*).—4, *Fagraea ceilanica*-type, *Fagraea annulata* subtype (*F. eymae*).—5-6, *Fagraea fragrans*-type, *Fagraea fragrans* subtype (*F. fragrans*).—7-9, *Fagraea fragrans*-type, *Fagraea racemosa* subtype (*F. gracilipes*).—10-11, *Fagraea berteriana*-type (*F. salticola*). $\times 1100$.



Pl. 7. Replicas. 1 (*Fagraea ceilanica*). $\times 2800$.—2 (*Fagraea blumei*). $\times 5200$.

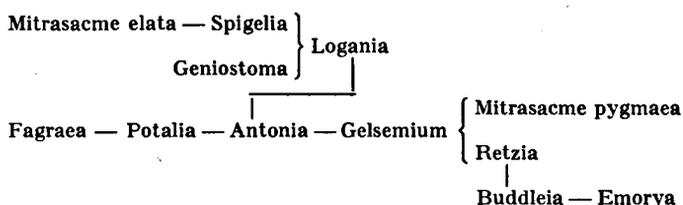
A Systematic Commentary

(P. W. Leenhouts)

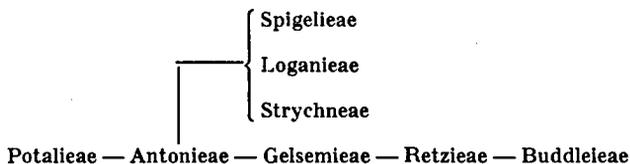
Dr. Punt intentionally chose for his systematic-palynological study a family on which in recent years much systematic work has been done or is still in progress. First of all this makes it possible to compare palynological results with the systematics as based upon the data derived from other disciplines, mainly gross morphology, and to have this done by a systematist specialized in that family. Furthermore, this had three more advantages for him: 1) he could be guided in selecting the taxa which deserved to be studied in the first place, 2) revised material was available, and 3) he could discuss his results with specialized taxonomists.

Before discussing the different palynological types in more detail, we may compare a scheme of their relationships as they are given in the preceding paper (Scheme I) with a scheme of the supposed relationships of the tribes based upon gross morphology (Scheme III). To facilitate the comparison, in Scheme II the first scheme has been "translated" into tribes.

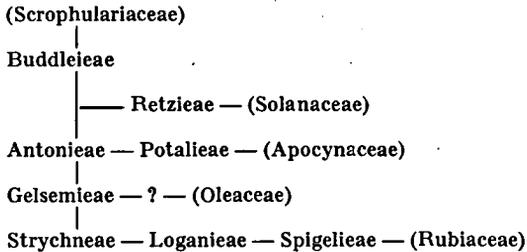
Scheme I. Relationships of palynological types.



Scheme II. Relationships of tribes based upon palynology.



Scheme III. Relationships of tribes based upon gross morphology.



When comparing the scheme of the relationships of the tribes as based upon gross morphology (Scheme III) with that based upon palynology (Scheme II) the overall similarity is striking. In both the basic group comprises the Antonieae and the Gelsemieae, the Potalieae are connected with the Antonieae, the series Strychneae—Loganieae—Spigeliae is connected more on the side of the Gelsemieae. The Retzieae connect in both schemes the Buddleieae more or less with the further Loganiaceae, though in Scheme II the Retzieae are placed more towards the Gelsemieae, in Scheme III more towards the Antonieae.

ANTONIA-TYPE

The mutual connexion of the four genera included in the tribe Antonieae—viz. *Bonyunia*, *Antonia*, *Norrisia*, and *Usteria*—is confirmed by palynological evidence. Whereas palynologically *Bonyunia* is slightly different, *Usteria* is gross morphologically more isolated, showing several specializations in the flower. The resemblance of this pollentype with several other types of the Loganiaceae is well in accordance with the central position of this tribe in the family.

BUDDLEIA-TYPE

The Buddleia-type of pollen characterizes the Buddleieae apart from the genus *Emorya*. This is the first point of systematic interest, a discussion of which will be given, however, under the Emorya-type. The second point which deserves more attention is the position of the Buddleieae in relationship to the Loganiaceae proper on the one side, to the Scrophulariaceae on the other. I have discussed this position at some length some years ago (Leenhouts 1962). The conclusion was that morphologically as well as anatomically the Buddleieae clearly take a position between Loganiaceae and Scrophu-

lariaceae. However, the delimitation against the Loganiaceae is more vague, its inclusion in that family seems more natural, than is the case in relation to the Scrophulariaceae. The genus *Peltanthera*, on the one end, shows distinct relationships with the Antonieae, *Buddleia* on the other side comes nearest to the Scrophulariaceae. For the systematist it is disappointing that palynology—like phytochemistry—clearly reveals a close relationship to the Scrophulariaceae and hardly any to the Loganiaceae. See, however, also under the *Retzia*-type.

The two subtypes are systematically unimportant.

EMORYA-TYPE

As alluded to already under the *Buddleia*-type, the fact that the genus *Emorya* is apparently characterized among all *Buddleieae* by a different—though related—pollentype is interesting. *Emorya* is a monotypic genus from southwestern North America. It is at least closely related to *Buddleia*, should possibly even be included in that genus judging from gross morphology. There is one palynological fact that may point in the same direction, however. Out of the several *Buddleia* species, studied by Punt—about half of which American—two (*B. alpina* and *B. americana*), both American species, show sometimes, resp. usually 4-colporate pollen, thus partly breaking down the differences between the two types. It is possible that if more American species of *Buddleia* would be studied palynologically, more of these cases could be found, and hence the demarcation between these two types would turn out to be less sharp.

SPIGELIA-TYPE

This type characterizes two out of four genera of the *Spigeliaeae*, viz. *Spigelia* and *Polypremum*, and the genus *Desfontainia* of uncertain position. According to Punt it is nearest related to the *Logania* type—to which among others belong *Cynoctonum* and part of *Mitrasacme*, the two other genera of the *Spigeliaeae*—and to the *Mitrasacme elata* type to which belongs another part of *Mitrasacme*. Palynological evidence hence distinctly supports the relationship between the *Spigeliaeae* and the *Loganieae* suggested in the third scheme though in detail there is a discrepancy between systematics and palynology.

Contrary to the great morphological resemblance between the

Spigeliaceae and the Rubiaceae-Hedyotideae, the pollen of these two tribes are completely different.

Palynological evidence now strengthens the argumentation for the inclusion of *Polypremum* in the Spigeliaceae instead of in the Buddleiaceae, as was usually done. Likewise, palynological evidence is in favour of the inclusion of *Desfontainia* in the Loganiaceae—though inclusion in the Spigeliaceae seems out of question.

MITRASACME-TYPES

From a palynological point of view, *Mitrasacme* is doubtless one of the most interesting genera among the Loganiaceae. Though only six out of a total of about 40 species have been studied, these represent already four distinct types. They are the *M. elata*-type (distinctly related to the Spigelia-type), the Logania-type, the *M. indica* type (close to the Logania-type), and the *M. pygmaea*-type (related to the Gelsemium-type). There is no doubt that *Mitrasacme* has to be included in the Spigeliaceae and that it represents an entity taxonomically. As the genus is in urgent need of revision, hardly anything can further be said. Only can be suggested that *M. elata* may belong to another subgenus or section than the other species studied.

LOGANIA-TYPE

The genera showing this type belong to three different, though doubtless mutually related, tribes: the Strychnaceae (complete), the Loganiaceae (*Logania*), and the Spigeliaceae (*Cynoctonum* and at least one species of *Mitrasacme*). The palynological resemblance with the *Mitrasacme indica*-type, the Spigelia-type, and the Geniostoma-type also reflect the closer connexion between these three tribes. Especially in this part of the family the palynological demarcations do not coincide with those based upon gross morphology.

GELSEMIUM-TYPE

Well in accordance with the conclusions derived from gross morphology, *Gelsemium* and *Mostuea* are also palynologically distinctly closely related. The resemblance with the Antonia- and Logania-types is also in accordance with the supposed systematic relationships.

RETZIA-TYPE

This type characterizes the monotypic tribe Retzieae. The position of the genus *Retzia* has been uncertain for a long time. Recently, Leeuwenberg (1964) included it in the Loganiaceae as a tribe of its own, related to both the Antonieae and the Buddleieae. It is very satisfying that palynological evidence favours its inclusion in the Loganiaceae, and even to some degree its exact place in this family. For the Retzia-type comes nearest to the Gelsemium-type—the Gelsemieae being distinctly related to the Antonieae—the Buddleia type following at the second place. Moreover, this strengthens the position of the Buddleieae within the Loganiaceae. On the other hand, the differences in pollen between *Retzia*, resp. *Metternichia* and *Sessea* are in contrast to a relationship with the Solanaceae as also often suggested.

GENIOSTOMA-TYPE

This type characterizes two out of the three genera of the Loganiaceae, *Logania*—which shows a certain similarity palynologically—being included under the Logania-type. These two genera are systematically intimately related; actually *Labordia* is hardly more than an isolated, in some characters more specialized offshoot of *Geniostoma*. The two subtypes are apparently hardly of any systematical importance.

POTALIA-TYPE

The Potalia-type and the Fagraea-types cover the genera *Fagraea*, *Potalia* and *Anthocleista* which together make up the tribe Potalieae. The Potalia-type is via *Fagraea gardenioides* and *F. berteriana* connected with the Fagraea berteriana-type which is the highest evolved one among the Fagraea-types. This position of the Potalia-type is in good accordance with the systematic affinities as *Fagraea* includes the doubtless most primitive members of the Potalieae, while *Potalia* and *Anthocleista* are clearly more specialized genera, especially as regards their flower characters.

The striking resemblance between Loganiaceae-Potalieae and Apocynaceae-Tabernaemontanae is not supported by palynology.

FAGRAEA-TYPES

Palynologically, *Mitrasacme* and *Fagraea* appear to be the most interesting genera of the Loganiaceae. Whereas *Mitrasacme* is

systematically as well as palynologically too insufficiently known to draw any conclusions, our knowledge of the genus *Fagraea* seems to justify a closer consideration. Twenty-five out of thirty-one species have been studied palynologically, and a recent taxonomical revision is available (Leenhouts 1962).

Parallel with the grouping into palynological types and subtypes and the "evolutionary trends" given by Punt in the main part of this paper, a grouping of the species on morphological grounds and some more "evolutionary trends" derived from gross morphology may precede a closer study of the systematic relationships.

The species of *Fagraea* can be grouped as follows:

elliptica, *fragrans* and probably *umbelliflora*, together making up the section *Cyrtophyllum*; possibly, there is some relationship between *umbelliflora* and *gracilipes*;

racemosa, the only species of the section *Racemosae*;

ceilanica, *annulata* (which shows also relationship to the *berteriana*-group), *acuminatissima*, *tubulosa*, *ridleyi*, *blumei*, *fastigiata*, possibly *crenulata*, *truncata*, *tacapala*, *woodiana*, *longiflora*, and *carstensensis*; *auriculata*, *involucrata*, *macroscypha*, and *resinosa*; this group is doubtless related to the previous one;

carnosa;

gardenioides, *curtisii*, and *calcareae*; this group is possibly related with the *berteriana*-group;

gracilipes (relationships possibly with *umbelliflora* and *berteriana*); *eymae*, *salticola*, *bodenii*, *gitingensis*, and *berteriana*; relationships possibly with *annulata*, the *gardenioides*-group, and *gracilipes*.

The "evolutionary trends" underlying the assumptions on primitive or advanced are:

1. tree → shrub or epiphyte; 2. leafbase without auricles → with auricles (wings are possibly a transitional stage phylogenetically);
3. inflorescences many-flowered → few-flowered → flowers solitary;
4. inflorescences dichasial → umbellate or glomerulous; 5. inflorescences dichasial → thyrsoid → racemose → spicate; 6. pedicels with one pair of bracteoles → without bracteoles; 7. pedicels with one pair of bracteoles → with two pairs of bracteoles; 8. bracteoles rather small and inserted halfway the pedicel → big, forming a kind

of involucre around the calyx. Other probable trends are: 9. inflorescences terminal → axillary; 10. stamens inserted directly on the corolla → inserted on a thickened ring; 11. anthers about oblong → linear; 12. stigma undivided → bilobed.

Now we may have a closer look at the different groups of species in the light of the "phylogenetical" series mentioned above and of palynological evidence.

Sect. *Cyrtophyllum* represents doubtless the most primitive group in *Fagraea*. *F. elliptica* is primitive according to 11 of the criteria mentioned above, only the bracteoles being occasionally absent. *F. fragrans* seems to be slightly more advanced (inflorescences axillary, sometimes few-flowered, bracteoles sometimes absent). *F. umbelliflora* is distinctly more advanced in several characters. Furthermore, these species fit better in with the whole of the Loganiaceae—especially with the Antonieae—than the other groups. *F. elliptica* and *fragrans* are both widespread—throughout Malesia, resp. also in continental Asia—which may also account for relative primitiveness; *F. umbelliflora* is a local endemic of New Guinea. All this is very well in accordance with palynological evidence: all three are characterised by *F. fragrans*-type pollen which is considered the most primitive type in the genus. This is especially satisfying as to *F. umbelliflora* as its inclusion in this section was provisional only. Moreover, its further relationship with *F. gracilipes* is also confirmed palynologically! The *fragrans* and *gracilipes* subtypes may be of less importance systematically.

F. racemosa, representing the monotypic section *Racemosae*, is, according to the trends cited above, relatively primitive. Only in its mostly shrublike habit and in the thyrsoid, racemose, spicate, or glomerulous inflorescences—in the last case rather few-flowered—is it more advanced. Its relatively wide geographical distribution—from SE. Asia to the Solomon Islands and northern Australia—also points to relative primitiveness. This is in accordance with palynology, as its pollen belong to the supposedly primitive *F. fragrans*-type.

The *ceilanica* group is the kernel of the section *Fagraea*. It seems too speculative and hardly of any use to try at a phylogenetic scheme within this group. Some of its species are about as primitive as *F. elliptica* and *fragrans* according to the above criteria, others show specializations mainly regarding shrublike or epiphytic habit, wings at the leafbase, few-flowered and glomerulous inflorescences, some-

times reductions or specializations in the bracteoles, in one case insertion of the stamens on a ring (*F. annulata*). First to be mentioned among the more primitive species are *F. crenulata* which will be discussed later, and *F. blumei* and, *F. tacapala* and *F. ceilanica*. *F. blumei* and *ceilanica*, both widespread, are doubtless closely related, and together constitute the base of the group. *F. ceilanica* is very variable, but can not be subdivided in well defined taxa; *F. blumei* is towards the east replaced by some taxa which show more advanced characters (ssp. *plumeriaeflora*, *F. fastigiata*, *truncata*, and *tacapala*). Palynologically, all species of this group have pollen of the *F. ceilanica*-type with the exception only of *F. crenulata* the pollen of which are included as *crenulata* subtype under the *F. fragrans*-type. *F. crenulata* is a rather primitive, relatively widespread species, which probably had better not be included in the *ceilanica* group. Furthermore, the subdivision of the *ceilanica*-type into two subtypes is of interest. Most of the species show the *ceilanica* subtype, *F. annulata* and *carstensensis* make up the *annulata* subtype. As a matter of fact, a palynological series can be made: *F. ceilanica*–*carstensensis*–*annulata*–*eymae*–*berteriana*. This series reflects surprisingly well the main systematic relationships. *F. carstensensis*, *annulata* and *eymae* are all rare local endemics of western New Guinea, where *F. ceilanica* has developed some related races. The position of *F. carstensensis* was morphologically not very clear; *F. annulata* is doubtless closely related to *F. ceilanica*, different, however, by the insertion of the stamens on a ring, a character pointing to the *berteriana* group; *F. eymae* and *berteriana* belong to the latter group and will be further discussed there. Here again the agreement between systematic conclusions derived from palynology, resp. gross morphology, is very satisfactory.

The *auriculata* group is a very coherent one. It consists of the widespread *F. auriculata*, and three Bornean endemics. The relationship seems to be with the *ceilanica* group, more especially with the alliance of *F. blumei*. The *auriculata* group is more advanced as a whole, however, by the tendency towards a solitary flower, towards two or three pairs of bracteoles, forming an involucre around the calyx, by the in principle epiphytic or shrublike habit, and by the auricles at the leafbase (missing in *F. resinosa*, possibly secondary?). Palynologically, all four species show the *ceilanica* subtype of pollen, thus stressing the relationship with the *ceilanica* group.

F. carnosa seems systematically rather isolated. Morphologically it is rather advanced in some characters (epiphyte, flowers solitary). It is relatively widespread (Lower Burma and West Malesia). Palynologically, it is included in the advanced berteriana type. There seems to be no reason, however, to look at the berteriana group—to which belong the other species with this type of pollen—for closer relationships.

The gardenioides group, distinctly cohering, shows no distinct closer relationships to any of the others. Only the presence of linear anthers and of a sometimes bilobed stigma may point to the berteriana group. This resemblance, up till now hardly taken serious—the gardenioides group being West Malesian till Borneo, the berteriana group East Malesian, though including the Philippines—is now strengthened by palynology. Only the pollen of *F. gardenioides* ssp. *borneensis* are known; they belong to the relatively advanced Potalia-type, show a distinct resemblance to *F. berteriana* pollen, however!

F. gracilipes is a rather isolated, in most characters relatively primitive species. Its area of distribution is fairly wide—from New Guinea and NE. Australia to Fiji; within this area about 5 local races are more or less distinguishable. This too speaks well for relative primitiveness. This is confirmed by its pollen which belong to the gracilipes subtype of the fragrans-type. Moreover, this strengthens the suggested relationship between *F. gracilipes* and *umbelliflora*.

The berteriana group is again very coherent. Apart from *F. eymae*—which seems to be related to *F. salticola*, however—they all show the probably advanced characters of a thickened ring on which the stamens are inserted (furthermore known only from *F. annulata*), linear anthers (furthermore in *F. gardenioides*), and a distinctly bilobed stigma (a tendency to which is also found in *F. woodiana*, *gardenioides*, and *gracilipes*). Furthermore, *F. berteriana* lacks bracteoles. Apart from *F. gitingensis* (Philippines and Moluccas) and *F. berteriana* (New Guinea, NE. Australia, and widely distributed in the SW. Pacific), it are endemics of New Guinea. *F. berteriana* shows the most advanced characters, and, though wide, its area of distribution makes also the impression of being young (for argumentation see Leenhouts 1962). These systematical reflections are fully supported by palynological evidence. All four species known (*F. gitingensis* is unknown) have *F. berteriana*-type pollen, and, as alluded to already under the ceilanica group, this type is connected

with the *F. ceilanica*-type via a series *F. ceilanica-carstensensis-annulata-eymae-berteriana*. In how far the close similarity between *F. berteriana* pollen and *F. gardenioides* pollen—the latter representing the most advanced *Potalia*-type—reflects real close relationship can hardly be said; as *F. gardenioides* shows also the linear anthers and a tendency towards the bilobed stigma of the *berteriana* group, and as *F. gardenioides* reaches to Borneo, the *berteriana* group to the Philippines, this is far from impossible. Then, however, *F. gardenioides* may represent a rather early offshoot.

Summarizing, the agreement between palynology and hitherto accepted systematics based nearly exclusively upon gross morphology is surprisingly good. Of course, this is a great satisfaction for the students of both disciplines. The fact that (in *Fagraea*) even detail problems in systematics could partly be solved thanks to palynological evidence, may be exceptional, but it also shows the value of studying the pollen of as many species of a genus as possible. This will only be possible, however, if somebody is working systematically in the taxon concerned or if a recent revision of the taxon is available.

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